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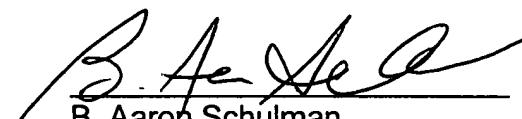
Assistant Commissioner of Patents

Washington, D. C.

SIR:

Applicant hereby claims the priority date of the attached under the provisions of 35 U.S.C. 119.

Respectfully submitted,



B. Aaron Schulman  
Registration No. 31877

Date: 28 June 2004

STITES & HARBISON PLLC  
Transpotomac Plaza  
1199 North Fairfax Street, Suite 900  
Alexandria, Virginia 22314  
(703) 739-4900



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Attestation

Die angehefteten Unterlagen stimmen mit der ursprünglich eingereichten Fassung der auf dem nächsten Blatt bezeichneten europäischen Patentanmeldung überein.

The attached documents are exact copies of the European patent application described on the following page, as originally filed.

Les documents fixés à cette attestation sont conformes à la version initialement déposée de la demande de brevet européen spécifiée à la page suivante.

Patentanmeldung Nr. Patent application No. Demande de brevet n°

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Der Präsident des Europäischen Patentamts;  
Im Auftrag

For the President of the European Patent Office

Le Président de l'Office européen des brevets  
p.o.

R C van Dijk

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Anmelder/Applicant(s)/Demandeur(s):

INSTITUT NATIONAL DE LA SANTE ET DE LA  
RECHERCHE MEDICALE (INSERM)  
101, rue de Tolbiac  
75013 Paris  
FRANCE

Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:  
(Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung.  
If no title is shown please refer to the description.  
Si aucun titre n'est indiqué se referer à la description.)

Infectious pestivirus pseudo-particles containing functional envelope proteins, E1, E2  
envelope proteins

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Infectious pestivirus pseudo-particles containing functional  
Erns, E1, E2 envelope proteins

5 The invention relates to the generation and the use of pestivirus pseudo-particles containing native functional E1, E2 envelope glycoproteins assembled onto retroviral core particles. These particles are highly infectious and constitute a valid model of pestivirus virion.

10 Pestivirus are single-stranded RNA (ssRNA) enveloped spherical viruses that constitute a genus within the family Flaviviridae, which also includes the genera 15 flavivirus and hepacivirus (human hepatitis C viruses). Several pestiviruses are important mammalian pathogens, especially cattle pathogens, such as the bovine viral diarrhea, the swine fever and the border disease viruses. Pestivirus can cause mucosal diseases (diarrhea), respiratory disease, suppression of an animal's immune system, and severe bleeding disorders.

15 Pestivirus structural proteins and non structural proteins are expressed from a single polyprotein precursor and individually released in their respective cell compartments upon cleavage by cellular and viral proteases. By analogy with other members of the Flaviviridae, pestivirus genomic organization suggests a virus 20 consisting of a nucleocapsid comprising a viral genome and core protein (C) coated by a lipid envelop containing the two envelope glycoproteins E1 and E2.

25 The majority of acute bovine viral diarrhea virus (BVDV) infections are caused by noncytopathic viruses. Cattle acutely or persistently infected with BVDV are the primary source of virus. Infected animals shed virus in nasal and oral secretions, feces and urine. The primary virus entrance route is probably oral nasally. Other less 30 important routes of entry may include infected semen, biting insects, and contaminated instruments. Following entry and contact with the mucosal lining of the mouth or nose, initial replication occurs in epithelial cells with a predilection for the palatine tonsils. From here, the virus is able to spread systemically through the blood stream. Spread can occur through both free virus in the serum and virus infected leucocytes, particularly lymphocytes and monocytes. Isolation of virus from serum or leucocytes is generally possible between 3 and 10 days post infection. During systemic spread, the virus is able to gain entry to most tissues with a preference for lymphoid tissues. BVDV broadly infects cattle, sheep, goats, and pigs.

Classical swine fever disease (SFV, previously called hog cholera virus) is another member of the family Flaviviridae, genus Pestivirus. SFV is an economically important contagious disease of swine world-wide. The disease occurs in much of Asia, Central and South America, and parts of Europe and Africa. Several countries

5 have eradication programs in force, based on rapid diagnosis and stamping out of infected herds, supplemented by other control measures. Despite these efforts, SFV has still not been eliminated in many countries. Although SFV can replicate in non-porcine cells, porcine kidney cells are used most frequently for virus growth. Virus replication is restricted to the cytoplasm of the cell and does not result in a cytopathic

10 effect. The first progeny virus is released from the cells at 5-6 hours post-infection. Virion assembly occurs on membranes of the endoplasmic reticulum, but performed capsids and budding are not seen. Instead, fully formed virions appear within the cisternae of the endoplasmic reticulum and are released via exocytosis or cell lysis. Pigs and wild boar are the natural hosts of SFV.

15 Border disease (BD) is a congenital disease of sheep that was first reported in the bordering countries of England and Wales. A similar, but rare condition also occurs in goats. The causative agent of BD, the border disease virus (BDV), is found worldwide in sheep. Five to fifty percent of sheep tested have antibodies against BD virus; meaning that these ewes have either been exposed to, or are carrying the

20 disease. Transmission of the virus occurs via oral and/or intranasal routes in sheep. Persistently infected sheep are the primary virus reservoir. These ewes will shed virus in all excretions and secretions. Lambs of persistently infected ewes are at risk of becoming persistently infected with the BDV, and thereby perpetuating the disease cycle.

25 The invention describes the formation and use of infectious pestivirus pseudo-particles harboring unmodified E1 and E2 glycoproteins.

#### *Definitions*

The terms "vector", "cloning vector" and "expression vector" mean the vehicle

30 by which a DNA or RNA sequence (e.g. a foreign gene) can be introduced into a host cell, so as to transform the host and promote expression (e.g. transcription and translation) of the introduced sequence. Vectors typically comprise the DNA of a transmissible agent, into which foreign DNA is inserted. A common way to insert one segment of DNA into another segment of DNA involves the use of enzymes called

restriction enzymes that cleave DNA at specific sites (specific groups of nucleotides) called restriction sites. Generally, foreign DNA is inserted at one or more restriction sites of the vector DNA, and then is carried by the vector into a host cell along with the transmissible vector DNA. A segment or sequence of DNA having inserted or 5 added DNA, such as an expression vector, can also be called a "*DNA construct*". A common type of vector is a "*plasmid*", which generally is a self-contained molecule of double-stranded DNA, usually of bacterial origin, that can readily accept additional (foreign) DNA and which can readily be introduced into a suitable host cell. A plasmid 10 vector often contains coding DNA and promoter DNA and has one or more restriction sites suitable for inserting foreign DNA. Coding DNA is a DNA sequence that encodes a particular amino acid sequence for a particular protein or enzyme. Promoter DNA is a DNA sequence that initiates, regulates, or otherwise mediates or 15 controls the expression of the coding DNA. Promoter DNA and coding DNA may be from the same gene or from different genes, and may be from the same or different organisms. A large number of vectors, including plasmid and fungal vectors, have 20 been described for replication and/or expression in a variety of eukaryotic and prokaryotic hosts.

A "*coding sequence*" or a sequence "*encoding*" an expression product, such as a RNA, polypeptide, protein, or enzyme, is a nucleotide sequence that, when 25 expressed, results in the production of that RNA, polypeptide, protein, or enzyme, i.e., the nucleotide sequence encodes an amino acid sequence for that polypeptide, protein or enzyme.

The term "*transfection*" means the introduction of a foreign nucleic acid (DNA, cDNA or RNA) into a cell so that the host cell will express the introduced gene or 30 sequence to produce a desired substance, typically a protein coded by the introduced gene or sequence. The introduced gene may include regulatory or control sequences, such as start, stop, promoter, signal, secretion, or other sequences used by a cell's genetic machinery. A host cell that receives and expresses introduced DNA or RNA has been "*transformed*".

The term "*host cell*" means any cell of any organism that is selected, modified, transformed, grown, or used or manipulated in any way, for the production of a substance by the cell, for example the expression by the cell of a gene, a DNA sequence, a protein, a virion. In the context of the invention, the host cell is a mammalian cell, preferably a cell from cattle, rabbit, pig, goat, swine. Suitable host

cells include for instance epithelial cells, leucocytes, lymphocytes, macrophages, monocytes, primary kidney cells from cattle or pig, and BT cells (ATCC CRL-1390).

As used herein, the term "*permissive cell*" is meant for a cell that is permissive for a pestivirus infection.

5 "Pestiviruses" are members of the *Flaviviridae* family. Pestivirus genome encodes a single polyprotein NH<sub>2</sub>-C-Erns-E1-E2-p7-NS2-NS3-NS4a-NS4b-NS5a-NS5b-COOH that is processed co and post-translationally into both structural (N-terminal nucleocapsid protein termed "Core" (C), and proteins Erns, E1 and E2) and non-structural (NS) proteins. The amino-terminal part of the polyprotein is cleaved by 10 host cell proteases and its products, core and envelope (Erns, E1 and E2) proteins, are believed to be the major constituents of pestivirus particles (virions). However, the ectodomain Erns-E1 is thought to be processed upon synthesis, thus releasing the non anchored Erns protein.

15 Although most cleavages in the polyprotein precursor proceed to completion during or immediately after translation, processing between E2 and p7, a hydrophobic domain found at the carboxy terminus of E2, is incomplete and results in the production of fully processed E2 and uncleaved E2-p7.

20 In the context of the invention, said pestivirus may be of any specie, genotype, subtype, or variant of pestivirus strains. Preferably, the pestivirus according to the invention is selected from the group consisting of bovine viral diarrhea virus (BVDV), Type I or Type II, swine fever virus (SFV) and border disease virus (BDV). The complete genome sequence of BVDV (Genebank : NC\_001461), SFV (Genbank : NC\_002657) and BDV (Genbank : NC\_003679) is shown in SEQ ID No 1, 7 and 13, respectively.

25 The term "variant" refers to the homologous polynucleotide sequences and corresponding amino acid sequences found in the different pestivirus strains owing to pestivirus hypervariability.

The term "*pestivirus-like particles*" as used herein refers to non naturally occurring viral particles that comprise an envelope protein of an pestivirus.

30 The pestivirus pseudo-particles of the invention are infectious for a target cell. The particles of the invention more particularly comprise retroviral core proteins. Such particles may be readily produced by one skilled in genetic engineering techniques. One can for instance refer to EP 1 201 750 that describes production of

synthetic retroviral particles expressing an antigen for modulating an immune response.

In the context of the invention, the term "*infectious*" is used to describe the capacity of the particles of the invention to complete the initial steps of viral cycle that 5 lead to cell entry. However, upon interaction with the host cell, pestivirus-like particles may or may not produce progeny viruses.

The term "*an envelope protein of a pestivirus*" denotes the native Erns, E1 or E2 glycoprotein of a pestivirus, or a mutant thereof.

By an "*Erns glycoprotein*" or "*Erns protein*" is meant a Erns from any specie, 10 genotype, subtype, or variant of perstivirus strains. The amino acid sequence of BVDV, SFV and BDV Erns protein is shown in SEQ ID No 3, 9, and 15, respectively.

By an "*E1 glycoprotein*" or "*E1 protein*" is meant a envelope 1 protein (E1) from any specie, genotype, subtype, or variant of perstivirus strains. The amino acid sequence of BVDV, SFV and BDV E1 protein is shown in SEQ ID No 4, 10, and 16, 15 respectively.

By an "*E2 glycoprotein*" or "*E2 protein*" is meant a envelope 2 protein (E2) from any specie, genotype, subtype, or variant of perstivirus strains. The amino acid sequence of BVDV, SFV and BDV E2 protein is shown in SEQ ID No 5, 11, and 17, respectively.

20 By a "*p7 protein*" is meant a native pestivirus p7 protein, or a mutant thereof, from any specie, genotype, subtype, or variant of perstivirus strains. The amino acid sequence of BVDV, SFV and BDV p7 protein is shown in SEQ ID No 6, 12, and 18, respectively.

25 Preferably, Erns, E1, E2, and p7 glycoproteins are derived from a same pestivirus strain. Preferably said Erns and/or E1 and/or E2 and/or p7 proteins are native pestivirus proteins.

The term "*mutant*" or "*mutation*" is meant for alteration of the DNA sequence that result in a modification of the amino acid sequence of native Erns, E1, E2, or p7 30 proteins. Such a modification can be for instance the substitution and/or deletion of one or more amino acids. Mutants notably include fragments of native Erns, E1, E2 and p7 proteins. Variants are particular examples of naturally occurring mutants. Mutants are more particularly contemplated as useful for identifying the structural elements of Erns and/or E1 and/or E2 proteins, and optionally p7 protein, necessary for maintaining cell infectivity or for increasing Erns and/or E1 and/or E2 antigenicity

for vaccination purposes. In a preferred embodiment, the mutants encompass E2 glycoproteins wherein hypervariable region I has been deleted, while the particles produced therefrom remain infectious.

5 The term "*pestivirus core*" is meant for a native core protein of a pestivirus strains, a fragment thereof, or a variant thereof from any specie, genotype, subtype, or variant of pestivirus strains. According to an embodiment, the core protein is a N-terminally truncated form of pestivirus core ( $\Delta C$ ) that comprises the core signal peptide. The amino acid sequence of BVDV, SFV and BDV core protein is shown in SEQ ID No 2, 8, and 14, respectively.

10 The term "*polyprotein*" as used herein is used to describe a protein construct made up of individual proteins that are joined together in a sequence whereby they retain their original relevant biological activities.

15 The term "*a polyprotein comprising a pestivirus core protein linked to pestivirus Erns and/or a pestivirus E1 protein and/or pestivirus E2 protein*", or "*a polyprotein comprising successively a pestivirus core protein, and a pestivirus Erns and/or a pestivirus E1 protein and/or pestivirus E2 protein*", includes the CE<sub>ns</sub>E1E2, CE2Er<sub>ns</sub>E1, CE<sub>ns</sub>E1, CE1E2, CE2E1, CE1, CE2,  $\Delta$ CE<sub>ns</sub>E1E2,  $\Delta$ CE2Er<sub>ns</sub>E1,  $\Delta$ CE<sub>ns</sub>E1,  $\Delta$ CE1E2,  $\Delta$ CE2E1,  $\Delta$ CE1, and  $\Delta$ CE2 polyproteins.

20 Optionally, said polyprotein further contain the p7 protein. The polyprotein comprising a pestivirus core protein linked to pestivirus Erns and/or E1 protein and/or pestivirus E2 protein thus additionally includes the CE<sub>ns</sub>E1E2p7, CE2Er<sub>ns</sub>E1p7, CE2p7Er<sub>ns</sub>E1, CE<sub>ns</sub>E1p7, CE1E2p7, CE2E1p7, CE1p7, CE2p7,  $\Delta$ CE<sub>ns</sub>E1E2p7,  $\Delta$ CE2Er<sub>ns</sub>E1p7,  $\Delta$ CE2p7Er<sub>ns</sub>E1,  $\Delta$ CE<sub>ns</sub>E1p7,  $\Delta$ CE1E2p7,  $\Delta$ CE2E1p7,  $\Delta$ CE2p7E1,  $\Delta$ CE1p7, and  $\Delta$ CE2p7 polyproteins.

25 "CE<sub>ns</sub>E1E2" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein and a pestivirus E2 protein. "CE2Er<sub>ns</sub>E1" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, a pestivirus Erns protein and a pestivirus E1 protein. "CE<sub>ns</sub>E1" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus Erns protein, and a pestivirus E1 protein. "CE1E2" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E1 protein and a pestivirus E2 protein. "CE2E1" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein and a pestivirus E1 protein. "CE1E2" denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus

E1 protein and a pestivirus E2 protein. "CE1" denotes a polyprotein comprising a pestivirus core protein linked to a pestivirus E1 protein. "CE2" denotes a polyprotein comprising a pestivirus core protein linked to a pestivirus E2 protein.

" $\Delta CEmsE1E2$ " denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein and a pestivirus E2 protein. " $\Delta CE2EmsE1$ " denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, a pestivirus Erns protein and a pestivirus E1 protein. " $\Delta CEmsE1$ " denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus Erns protein, and a pestivirus E1 protein. " $\Delta CE1E2$ " denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, and pestivirus E1 and pestivirus E2 proteins. " $\Delta CE2E1$ " denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, and pestivirus E2 and pestivirus E1 proteins. " $\Delta CE1$ " denotes a polyprotein comprising a carboxy terminus of pestivirus core protein linked to a pestivirus E1 protein. " $\Delta CE2$ " denotes a polyprotein comprising a carboxy terminus of pestivirus core protein linked to a pestivirus E2 protein.  $\Delta CEmsE1E2$ ,  $\Delta CE1E2$ , as well as  $\Delta CE2$ , have been built by inserting a stop codon at the end of E2, whereas  $\Delta CE2EmsE1$ ,  $\Delta CEmsE1$ ,  $\Delta CE1$ ,  $\Delta CE2E1$  have been built by inserting a stop codon at the end of E1.

" $CEmsE1E2p7$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein, a pestivirus E2 protein, and a pestivirus p7 protein. " $CE2ErnsE1p7$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, a pestivirus Erns protein, a pestivirus E1 protein, and a pestivirus p7 protein. " $CE2p7ErnsE1$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, a pestivirus p7 protein, a pestivirus Erns protein, and a pestivirus E1 protein. " $CErsE1p7$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein, and a pestivirus p7 protein. " $CE1E2p7$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E1 protein, a pestivirus E2 protein, and a pestivirus p7 protein. " $CE2p7E1$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, a pestivirus p7 protein, and a pestivirus E2 protein. " $CE2E1p7$ " denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, a pestivirus E2 protein, and a pestivirus p7 protein. " $CE1p7$ "

denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E1 protein, and a pestivirus p7 protein. “ $\Delta CE2p7$ ” denotes a polyprotein comprising successively a pestivirus core protein, a pestivirus E2 protein, and a pestivirus p7 protein.

5 “ $\Delta CEmsE1E2p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein a pestivirus E2 protein, and a pestivirus p7 protein. “ $\Delta CE2EmsE1p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, a pestivirus Erns protein, a pestivirus E1 protein, and a pestivirus p7 protein. “ $\Delta CE2p7EmsE1$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, a pestivirus p7 protein, a pestivirus Erns protein, and a pestivirus E1 protein.

10 “ $\Delta CEmsE1p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus Erns protein, a pestivirus E1 protein, and a pestivirus p7 protein. “ $\Delta CE1E2p7$ ” denotes a polyprotein comprising a carboxy terminus of pestivirus core protein, a pestivirus E1 protein, a pestivirus E2 protein, and a pestivirus p7 protein. “ $\Delta CE2E1p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, a pestivirus E1 protein, and a pestivirus p7 protein. “ $\Delta CE2p7E1$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, a pestivirus p7 protein, and a pestivirus E1 protein. “ $\Delta CE1p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E1 protein, and a p7 protein. “ $\Delta CE2p7$ ” denotes a polyprotein comprising successively a carboxy terminus of pestivirus core protein, a pestivirus E2 protein, and a p7 protein.  $\Delta CEmsE1E2p7$ ,  $\Delta CE2EmsE1p7$ ,  $\Delta CEmsE1p7$ ,  $\Delta CE1E2p7$ ,  $\Delta CE2E1p7$ ,  $\Delta CE1p7$  as well as  $\Delta CE2p7$ , have been built by inserting a stop codon at the end of p7 whereas  $\Delta CE2p7EmsE1$ , and  $\Delta CE2p7E1$  have been built by inserting a stop codon at the end of E1.

30 By “retrovirus” is meant a virus whose genome consists of a RNA molecule and that comprises a reverse-transcriptase, *i.e.* a member of the Retroviridae family. Retroviruses are divided into Oncovirus, Lentivirus and Spumavirus. Preferably said retrovirus is an oncovirus, *e.g.* MLV, ALV, RSV, or MPMV, a lentivirus, *e.g.* HIV-1, HIV-2, SIV, EIAV, or CAEV, or a spumavirus such as HFV. Genomes of these retroviruses are readily available in databanks.

In the context of the invention "*a nucleic sequence comprising a packaging competent retrovirus-derived genome*" is intended for a sequence that comprises the retroviral nucleic acid sequences known as "*cis*-acting" sequences. These include the Long Terminal Repeats (LTRs) for the control of transcription and integration, the psi sequence necessary for encapsidation, and the Primer Binding site (PBS) and polypurine track (PPT) sequences necessary for reverse transcription of the retroviral genome. Advantageously, said nucleic acid sequence comprising a packaging competent retrovirus-derived genome further comprises a transgene.

Said retroviral genome may be replication-defective or replication-competent, in the absence of any trans-complementing function. A replication-competent genome would further comprise the gag, pol, and env retroviral genes. In a replication-defective genome, the viral genes gag, pol, and env are deleted. However, assembly of viral pseudo-particles may be achieved by providing another vector that comprises gag, pol and env but that is defective for the "*cis*" sequences. Their expression allows the encapsidation of the transgene, excluding the genes necessary for the multiplication of the viral genome and for the formation of complete viral particles.

As used herein, the term "*transgene*" designates the gene that is expressed in the target cell upon infection by the particles of the invention.

Examples of transgenes include a gene encoding a molecule of therapeutic interest, a marker gene, a gene coding for an immune modulator, an antigen, or a suicide gene.

A "*marker gene*" denotes a gene whose expression is detectable. For instance marker gene expression can generate a detectable signal, such as a fluorescence emission, a chromogenic reaction, or confer a growth advantage to the cells wherein it is expressed (antibiotic resistance genes).

An "*immune modulator*" refers to the product of a gene that modifies the activity of the immune system of a subject *in vivo*. Examples of immune modulators include cytokines, (e.g. interleukins, interferons, or haematopoietic colony stimulating factors), chemokines, and the like. Expression of an immune modulator by transformed cells may change the cellular environment and alter differentiation of immune cells and thus modify the type and the strength of immune response elicited against a given antigen.

An "antigen" refers to a molecule, such as a peptide, a polypeptide or a protein, against which an immune response is sought. Said antigen may be for instance a tumor, a bacterial, a pathogenic, a proteic, or a viral antigen.

5 A "suicide gene" is meant for a gene whose expression in cells induces programmed-cell death (apoptosis) such as the conditional Herpes Simplex virus type I thymidine kinase gene.

10 The "core protein from a retrovirus" refers to proteins encoded by the gag and pol genes. The gag gene encodes a polyprotein which is further processed by the retroviral protease into structural proteins that comprise the core. The pol gene encodes the retroviral protease, reverse-transcriptase, and integrase.

15 A "pharmaceutically acceptable carrier" refers to any vehicle wherein the vaccine composition according to the invention may be formulated. It includes a saline solution such as phosphate buffer saline. In general, a diluent or carrier is selected on the basis of the mode and route of administration, and standard pharmaceutical practice.

In the context of the present application, "vaccination" is intended for prophylactic or therapeutical vaccination. "Therapeutical vaccination" is meant for vaccination of a patient with a pestivirus infection.

20 According to the invention, the term "subject" or "patient" is meant for any mammal likely to be infected with pestivirus. Cattle, sheep, pigs and goats are examples of hosts for pestiviruses,

#### *Production of pestivirus pseudo- particles*

25 The inventors have generated infectious pseudo-particles that contain functional, and more particularly unmodified, pestivirus glycoproteins assembled onto retroviral core particles. Pestivirus ErnsE1E2, and optionally p7, are expressed from a polyprotein containing the core (C) protein or a fragment thereof, in particular the carboxy-terminus of the C protein, which served as signal peptide for Erns and/or E1 and/or E2 glycoproteins.

30 The invention thus provides a method for producing pestivirus-like particles *ex vivo* comprising the steps of:

- providing a first nucleic acid sequence comprising a packaging competent retrovirus-derived genome;

- providing a second nucleic acid sequence comprising a cDNA encoding the core proteins from said retrovirus;
- providing a third nucleic acid sequence comprising a cDNA encoding a polyprotein comprising successively a pestivirus core protein, and a pestivirus E1s and/or a pestivirus E1 protein and/or a pestivirus E2 protein;

5 - transfecting host cells with said nucleic acid sequences and maintaining the transfected cells in culture for sufficient time to allow expression of the cDNAs to produce structural proteins from pestivirus and retrovirus; and allowing the structural proteins to form virus-like particles.

10 The invention further provides a method for producing pestivirus-like particles *in vivo*, which method comprises the steps of :

- providing a first nucleic acid sequence comprising a packaging competent retrovirus-derived genome;
- providing a second nucleic acid sequence comprising a cDNA encoding the core proteins from said retrovirus;
- providing a third nucleic acid sequence comprising a cDNA encoding a polyprotein comprising successively a pestivirus core protein, and a pestivirus E1s and/or a pestivirus E1 protein and/or a pestivirus E2 protein;
- transfecting cells of a subject *in vivo* with said nucleic acid sequences, to allow expression of the cDNAs to produce structural proteins from pestivirus and retrovirus; and to allow the structural proteins to form virus-like particles.

20 Another aspect of the invention is the use of three nucleic acid sequences for the preparation of a medicament useful as a vaccine against an pestivirus infection wherein the nucleic acid sequences are :

25 - a first nucleic acid sequence comprising a packaging competent retroviral-derived genome;

- a second nucleic acid sequence comprising a cDNA encoding core proteins from said retrovirus;
- a third nucleic acid sequence comprising a cDNA encoding a polyprotein comprising successively a pestivirus core protein, and a pestivirus E1s and/or a pestivirus E1 protein and/or a pestivirus E2 protein ;

30 and, when transferred into cells of a subject, the nucleic acid sequences allow the production of structural proteins from pestivirus and retrovirus, wherein the structural proteins form virus-like particles that are immunogenic.

For the purpose of transfection, said first, second and third nucleic acid sequences may be carried on a same vector, or on two or three separated vectors.

In particular, plasmoviruses, adenoretroviruses and replicating pseudo-viruses are examples of vectors suitable for carrying the above-mentioned sequences. A 5 plasmovirus vaccine consists in such a plasmid DNA preparation, that allow expression of pestivirus pseudo-particles after administration in an patient in order to elicit a immune response against said pestivirus. Administration of such a plasmovirus vaccine being achieved for preventive vaccination into people at risk for pestivirus-induced disease or for therapeutic vaccination into pestivirus-infected 10 patients. Adenoretroviruses consist in an alternative way to provide the above-mentioned nucleic acid sequences encoding pestivirus pseudo-particles. In this case, it is possible to design three independent adenoretroviruses, *i.e.* recombinant adenoviruses, that encode the three nucleic acid sequences mentioned above (retroviral core and genome and pestivirus glycoproteins), or, alternatively, it is also 15 possible to design a single adenoretrovirus, derived from "guttless" recombinant adenoviruses, that contains the different nucleic acid sequences. Such adenoretroviruses can be administered to patient as for plasmoviruses, in order to elicit an anti-pestivirus immune response. Replicating pseudo-retroviruses are another alternative possibility to express all the above-mentioned nucleic acid 20 sequences encoding the pestivirus pseudo-particles. Such structures are in fact pestivirus-pseudo-particles whose genome is engineered to allow, following infection, its propagation into cells of an inoculated patient, thereby inducing the production of further replicating pestivirus pseudo-particles. In this case the genome of a retrovirus is modified so as to express the pestivirus E1E2 glycoproteins in place of the 25 retroviral Env gene (encoding the retroviral glycoproteins). The genes encoding the retroviral core proteins are left unchanged. Futhermore an additional gene, encoding a marker gene or an immunomodulator, for example, can be expressed from this genome.

According to a specific embodiment, said packaging competent retroviral 30 genome and core proteins are derived from a retrovirus selected from the group consisting of MLV, ALV, RSV, MPMV, HIV-1, HIV-2, SIV, EIAV, CAEV, and HFV.

Advantageously, the packaging competent retroviral genome further comprises a marker gene or an immune modulator.

In the method of the invention, said polyprotein may comprise CE<sub>n</sub>sE1E2, CE<sub>2</sub>Er<sub>n</sub>sE1, CE<sub>n</sub>sE1, CE1E2, CE2E1, CE1, CE2, ΔCE<sub>n</sub>sE1E2, ΔCE2Er<sub>n</sub>sE1, ΔCE<sub>n</sub>sE1, ΔCE1E2, ΔCE2E1, ΔCE1, or ΔCE2 polyproteins.

Preferably, said third nucleic acid sequence comprises a cDNA encoding a 5 polyprotein that further comprises a pestivirus p7 protein. Thus, preferably said polyprotein comprises successively a pestivirus core protein, and a pestivirus Er<sub>n</sub>s protein and/or a pestivirus E1 protein and/or a pestivirus E2 protein, and optionally a pestivirus p7 protein. The polyprotein comprising a pestivirus core protein linked to pestivirus Er<sub>n</sub>s and/or E1 protein and/or pestivirus E2 protein thus additionally 10 comprises the CE<sub>n</sub>sE1E2p7, CE<sub>2</sub>Er<sub>n</sub>sE1p7, CE2p7Er<sub>n</sub>sE1, CE<sub>n</sub>sE1p7, CE1E2p7, CE2E1p7, CE2p7E1, CE1p7, CE2p7, ΔCE<sub>n</sub>sE1E2p7, ΔCE2Er<sub>n</sub>sE1p7, ΔCE2p7Er<sub>n</sub>sE1, ΔCE<sub>n</sub>sE1p7, ΔCE1E2p7, CE2E1p7, CE2p7E1, ΔCE1p7, and ΔCE2p7 polyproteins.

According to an embodiment, Er<sub>n</sub>s and/or E1 and/or E2, and optionally p7 15 protein, are native proteins. According to another embodiment, Er<sub>n</sub>s and/or E1 and/or E2 proteins, and optionally p7 protein, are mutated to obtain particles that are useful for characterizing the glycoprotein determinants for pestivirus infectivity.

Preferably, said Er<sub>n</sub>s, E1, E2, and optionally p7 proteins are derived from a same pestivirus strain.

20 According to another embodiment said pestivirus core protein is a carboxy terminus form (ΔC) of pestivirus core protein that comprises the core protein signal peptide .

Preferably said pestivirus is selected from the group consisting of bovine viral diarrhea virus (BVDV), swine fever virus (SFV), and Border disease virus (BDV).

25 In accordance with the present invention there may be employed conventional molecular biology, microbiology, and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook et al., 1989 ; DNA Cloning: A Practical Approach, Volumes I and II (D.N. Glover ed. 1985) ; Oligonucleotide Synthesis (M.J. Gait ed. 1984) ; Nucleic Acid Hybridization 30 [B.D. Hames & S.J. Higgins eds. (1985)] ; Transcription and Translation [B.D. Hames & S.J. Higgins, eds. (1984)] ; Animal Cell Culture [R.I. Freshney, ed. (1986)] ; Immobilized Cells and Enzymes [IRL Press, (1986)] ; B. Perbal, A Practical Guide To Molecular Cloning (1984) ; F.M. Ausubel et al., 1994.

In particular, the vectors of the invention may be introduced into the target cell by means of any technique known for the delivery of nucleic acids to the nucleus of cells, either in culture, *ex vivo*, or *in vivo*.

Introduction of the nucleic acid sequences may be performed by any 5 standard method well known by one skilled in the art, e.g. transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, or use of a gene gun (see for instance Wu et al., 1992 ; Wu et al, 1988).

10 The donor nucleic acid targeting system can also be introduced by lipofection. In certain embodiments, the use of liposomes and/or nanoparticles is contemplated for the introduction of the donor nucleic acid targeting system into host cells. Nanocapsules can generally entrap compounds in a stable and reproducible way. Ultrafine particles (sized around 0.1  $\mu\text{m}$ ) that can be designed using 15 biodegradable polyalkyl-cyanoacrylate polymers are contemplated for use in the present invention, and such particles may be easily made.

Liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs)). MLVs generally have diameters of from 25 nm to 4  $\mu\text{m}$ . Sonication of MLVs results in the formation of small unilamellar vesicles 20 (SUVs) with diameters in the range of 200 to 500  $\text{\AA}$ , containing an aqueous solution in the core. The use of cationic lipids may promote encapsulation of negatively charged nucleic acids, and also promote fusion with negatively charged cell membranes (Felgner et al., 1989).

25 *In vivo* targeted gene delivery is described in international patent publication WO 95/28 494. Alternatively, the vector can be introduced *in vivo* by lipofection, using liposomes or nanoparticles as above described. It is also possible to introduce the vector *in vivo* using techniques that are similar to the techniques that are employed *in vitro* (e.g. transfection, electroporation...).

30

#### *Transformed cells*

The invention further relates to a transformed host cell that contains :

- a first nucleic acid sequence comprising a packaging competent retrovirus-derived genome;

- a second nucleic acid sequence comprising a cDNA encoding the core proteins from said retrovirus; and
- a third nucleic acid sequence comprising a cDNA encoding a polyprotein comprising successively a pestivirus core protein, and a pestivirus Erns protein and/or a pestivirus E1 protein and/or a pestivirus E2 protein.

5

Preferably, said third nucleic acid sequence comprises a cDNA encoding a polyprotein that further comprises a pestivirus p7 protein. Thus, preferably said polyprotein comprises successively a pestivirus core protein, a pestivirus Erns protein and/or a pestivirus E1 protein and/or a pestivirus E2 protein, and optionally a 10 pestivirus p7 protein.

Such a transformed host cell is obtainable as described in a method above.

In another aspect, the invention relates to the use of a transformed host cell as defined above, for the identification of molecules capable of interfering with pestivirus entry in cells. The invention provides in particular a method of *ex vivo* screening or 15 identification of molecules capable of interfering with pestivirus entry in cells comprising comparison of the level of transformed host cell fusion to a target host cell, in the presence or the absence of a candidate molecule. Said method preferably comprises the steps consisting of:

- co-culturing a transformed host cell with a target host cell, in the absence or 20 presence of a candidate molecule, under conditions that allow syncytia formation, *i.e.* cell-cell fusion, and pestivirus-like particle entry in target host cell in the absence of any candidate molecule;
- assessing syncytia formation in the absence and in the presence of said candidate molecule;
- 25 - comparing syncytia formation measured in presence of said candidate molecule with syncytia formation measured in absence of any candidate molecule;
- identifying as a molecule capable of interfering with pestivirus entry the candidate molecule for which syncytia formation, as measured in the presence of said molecule, is decreased as compared to syncytia formation measured in the 30 absence of any candidate molecule.

Contacting a transformed host cell with a target host cell, and a candidate molecule can be carried out by contacting simultaneously said transformed host cell, target host cell and candidate molecule. Otherwise, two of these three elements can

be contacted under conditions sufficient to allow their interaction before addition of the third missing element.

Preferably said target host cell is not transformed, *i.e.* said target host cell does not contain at least one of the first, second, and third nucleic acid sequence as 5 defined above.

Syncytia formation can be readily assessed by one skilled in the art. Briefly, the coculture is submitted to a acidic pH drop by incubation for 5 min at pH-5 and incubated in a normal medium for an additional 12 hrs. Cultures are then stained by adding the May-Grunwald and Giemsa solutions (MERCK) according to the 10 manufacturer recommendations. Cells containing two or more nuclei can be defined as syncytia. A fusion index is then defined as the percentage of (N-S)/T where N is the number of nuclei in the syncytia, S is the number of syncytia and T is the total number of nuclei counted.

15 *Pestivirus-like particles*

In the method described above no structural modifications of the E1E2 glycoproteins are required for their correct assembly on retroviral cores. The method of the invention thus makes it possible to generate high titre infectious pestivirus pseudo-particles with functional E1E2 proteins. As demonstrated herein, these 20 particles constitute a valid model of pestivirus virions as regards to early steps of viral infection cycle.

The invention further relates to an infectious pestivirus-like particle, comprising the core proteins from a retrovirus, and Erns and/or E1 and/or E2 pestivirus glycoprotein(s), and optionally p7 protein. Such a particle is obtainable by a method 25 as described above.

According to an embodiment, the infectious particle of the invention may comprise native pestivirus E1 protein, or native pestivirus E2 protein, or native pestivirus Erns protein and native pestivirus E1 protein, or native pestivirus E1 protein and native pestivirus E2 protein, or native pestivirus Erns protein and native 30 pestivirus E1 protein and native pestivirus E2 protein. Preferably said Erns and E1, or E1 and E2, or Erns, E1 and E2 proteins are derived from a same pestivirus strain. According to another embodiment, Erns and/or E1 and/or E2 glycoproteins are mutated.

Preferably the above described infectious particle of the invention further comprise a native pestivirus p7 protein. Preferably, said E1 and E2 glycoproteins, and p7 protein are derived from a same pestivirus strain. Still preferably said Erns, E1 and E2 glycoproteins, and p7 protein are derived from a same pestivirus strain

5 According to another embodiment, Erns and/or E1 and/or E2 glycoproteins and/or p7 protein are mutated.

Preferably said pestivirus is selected from the group consisting of bovine viral diarrhea virus (BVDV), swine fever virus (SFV), and Border disease virus (BDV).

Said retrovirus may be selected from the group consisting of MLV, ALV, RSV, 10 MPMV, HIV-1, HIV-2, SIV, EIAV, CAEV, and HFV.

Advantageously, said infectious particles further carry a transgene. For instance said transgene may be a marker gene which make it possible to follow-up cell infection by the infectious particles of the invention and can find application for instance in the identification of a cell receptor involved in pestivirus entry. Said 15 transgene can also be a gene encoding a molecule of therapeutic interest and/or a suicide gene.

*Use of the infectious pestivirus-like particles of the invention*

High infectivity of these particles makes it possible for the investigation of the 20 role of pestivirus Erns, E1 and E2 glycoproteins and their potential receptors in cell entry, pestivirus host-range and neutralisation by antibodies from pestivirus patient sera.

The invention therefore concerns the use of a pestivirus-like infectious particle as described above, for *ex vivo* identification of a cell receptor for pestivirus Erns 25 and/or E1 and/or E2 glycoprotein.

According to an embodiment, the invention provides a method for *ex vivo* identification of a receptor for pestivirus Erns and/or E1 and/or E2 glycoprotein comprising detection of the binding of said particle to a cell receptor. More specifically, the method may comprise the steps consisting of:

30 - contacting a cell susceptible to pestivirus infection with an infectious pestivirus-like particle of the invention, under conditions sufficient to allow specific binding of said particle to a receptor expressed at the surface of said cell;

- detecting binding of said particle to a receptor; and
- identifying said receptor.

A cell susceptible to a pestivirus infection, may be for instance a kidney primary cell, or cell line, from cattle, pig, or sheep.

Detection of particle binding to a receptor can be achieved according to classical procedures well known by one skilled in the art. For instance, this could 5 involve radioactive, enzyme or fluorescent labelling of the particles of the invention, and subsequent detection with an appropriate method. A number of fluorescent materials are known and can be utilized as labels. These include, for example, fluorescein, rhodamine, auramine, Texas Red. Enzyme labels consist in conjugation of an enzyme to a molecule of interest, e.g. a polypeptide, and can be detected by 10 any of colorimetric, spectrophotometric, or fluorospectrophotometric techniques. Flow cytometry analysis (FACS) together with labelled antibodies directed against E1 or E2 proteins harboured by the pseudo-particles of the invention is also appropriate.

According to another embodiment, the invention provides a method for *ex vivo* identifying a cell receptor for a pestivirus comprising the step consisting of:

15 - transfecting a cell which is not permissive for pestivirus infection with a nucleic acid sequence encoding a protein likely to be a receptor for pestivirus;  
- contacting said transformed cell with a pestivirus-like particle of the invention;  
- determining whether said transformed cell has become permissive or not for pestivirus infection; and  
20 - identifying as a cell receptor for a pestivirus said protein expressed by the transformed cell that has become permissive.

Determination of whether the transformed cell has become permissive for pestivirus infection can be readily achieved using the pestivirus-like particles of the invention. In particular, where said particles carry a marker gene, such as GFP, 25 permissivity (*i.e.* the capacity of cells to be infected with a pestivirus, or with a pestivirus-like particle) can be assessed by FACS analysis of the transformed cells. Where the marker gene is an antibiotic resistance gene, identification of cells infected by the pestivirus-like particle is readily achieved through exposure to said antibiotic.

Where one does not suspect a given protein to be a receptor for pestivirus 30 entry, in cells, the above method can advantageously be adapted for the screening and the identification of a cell receptor for a pestivirus. In particular, an expression cDNA library can be prepared, for instance from a cDNA library obtained by reverse-transcription of cellular mRNAs from a cell permissive for pestivirus infection. Expression of such a cDNA library would be driven by a constitutive promoter whose

nucleic acid sequence has been fused to the cDNA library in suitable vectors. Such a library would contain a vector encoding a cell receptor for a pestivirus. Non permissive cells can then be transfected with this expression library and further screened for the identification of a cell receptor for a pestivirus.

5 To this end, the invention proposes a method for *ex vivo* identifying a cell receptor for pestivirus comprising the step consisting of:

- providing an expression cDNA library obtained from a cell permissive for pestivirus infection;

- transfecting cells that are not permissive for pestivirus infection with said expression cDNA library;

- contacting said transformed cells with pestivirus-like particles of the invention;

- identifying and isolating those transformed cells that have become permissive for pestivirus infection;

- isolating the expression vector transfected in cells that have become permissive; and

- identifying as a receptor for pestivirus the proteins encoded by the cDNA sequence of said isolated expression vectors.

Advantageously, the expression cDNA library is expressed from retroviral vectors that comprise glycoproteins that allow infection of the pestivirus non permissive cells. Such glycoproteins can be the VSV-G glycoprotein derived from vesicular stomatitis virus (VSV) whose receptor is expressed in most cell types *ex vivo*. Such viral particles can be assembled using a packaging competent retrovirus-derived genome that comprises the expression cDNA library, and optionally a marker gene. According to this embodiment the method for isolating the expression vector expressed in cells that have become permissive to infection by the pestivirus-like particles of the invention is greatly facilitated. Indeed this latter embodiment is particularly advantageous in that the process of cell infection with retroviral vectors has greater efficacy, as compared to cell transfection. Furthermore, cell infection leads to stable integration of viral genome in the cellular genome. Accordingly, transgenes, *i.e.* cDNA and marker gene that are carried by the pseudo-particles of the invention, are found to be stably expressed by infected cells. This in contrast with classical vectors used for transfection that do not integrate into cellular genome and for which expression may be transient.

In another aspect, the invention relates to the use of an infectious particle as defined above, for the identification of molecules capable of interfering with pestivirus entry in cells.

In particular, herein is provided a method of *ex vivo* screening or identification 5 of molecules capable of interfering with pestivirus entry in cells comprising comparison of the level of cell infection by the particles of the invention in the presence or the absence of a candidate molecule. Said method preferably comprises the steps consisting of:

- contacting a cell susceptible to pestivirus infection with an infectious pestivirus-like particle, in the absence or presence of a candidate molecule, under 10 conditions that allow cell infection with pestivirus-like particle in the absence of any candidate molecule;
- assessing cell infectivity in the absence and in the presence of said candidate molecule;
- comparing cell infectivity measured in presence of said candidate molecule 15 with cell infectivity measured in absence of any candidate molecule;
- identifying as a molecule capable of interfering with pestivirus entry the candidate molecule for which cell infectivity, as measured in the presence of said molecule, is decreased as compared to cell infectivity measured in the absence of 20 any candidate molecule.

Contacting a cell susceptible to pestivirus infection with an infectious pestivirus-like particle, and a candidate molecule can be carried out by contacting simultaneously said cell, pestivirus-like particle and candidate molecule. Otherwise, two of these three elements can be contacted under conditions sufficient to allow 25 their interaction before addition of the third missing element.

Cell infectivity can be readily assessed by one skilled in the art. One can take advantage of the embodiment wherein the infectious pestivirus-like particle carries a detectable marker gene to detect cell infection. In a preferred embodiment, the marker gene is a fluorescent marker gene, such as GFP, and the infection is 30 detected by means of fluorescence measurement, for instance by flow cytometry analysis of cells contacted with said infectious particles.

A cell suitable to be used in the method of identification of molecules interfering with pestivirus cell entry may be for instance a kidney primary cell, or cell line, from cattle, pig or sheep.

Such molecules capable of interfering with pestivirus entry in cells may constitute new antiviral drugs.

5 The infectious particles of the invention are further useful for diagnosis of pestivirus infection and follow-up of pestivirus infection, for instance to assess efficacy of a therapy in a patient.

The invention thus concerns the use of an infectious pestivirus-like particle for the *in vitro* detection of antibodies directed against pestivirus in a biological sample from a subject susceptible to be infected with pestivirus. Said biological sample may 10 be a biological fluid, such as blood or serum, or a tissue biopsy. In a specific embodiment, said antibodies are directed against E1 and/or E2 and/or E2 pestivirus proteins.

Accordingly, the invention provides a method of *in vitro* diagnosis of a pestivirus infection in a patient comprising detecting immune complexes formed by 15 interaction of anti-pestivirus antibodies likely to be present in a biological sample of the patient, with pestivirus-like particle of the invention. Said method may in particular comprise the steps consisting of:

- contacting a biological sample with an infectious pestivirus-like particle of the invention under conditions sufficient to allow formation of complexes by binding of 20 said infectious particle to antibodies directed against pestivirus present in the biological sample;
- detecting said complexes, which presence is indicative of a pestivirus infection.

The presence of antibodies reactive with pestivirus-like particles can be 25 detected using standard electrophoretic and immunodiagnostic techniques, including immunoassays such as competition, direct reaction, or sandwich type assays. Such assays include, but are not limited to, Western blots; agglutination tests; enzyme-labeled and mediated immunoassays, such as ELISAs; biotin/avidin type assays; radioimmunoassays; immunoelectrophoresis; immunoprecipitation, etc. The 30 reactions generally include revealing labels such as fluorescent, chemiluminescent, radioactive, enzymatic labels or dye molecules, or other methods for detecting the formation of a complex between the pestivirus-like particle and the antibody or antibodies reacted therewith.

In another embodiment, said method of *in vitro* diagnosis of a pestivirus infection in a patient comprises detecting an inhibitory effect of anti-pestivirus antibodies likely to be present in a biological sample of the patient, on the infection of a permissive cell by a pestivirus-like particle of the invention. Said method may in 5 particular comprise the steps consisting of:

- contacting a cell permissive for pestivirus infection with a pestivirus-like particle and a biological sample;
- comparing cell infectivity measured in presence of said biological sample with cell infectivity measured in absence of said biological sample;
- 10 - detecting the inhibition of pestivirus-like particle infection of a permissive cell as a decrease in cell infectivity measured in presence of said biological sample compared with cell infectivity measured in absence of said biological sample, said inhibition being indicative of a pestivirus infection.

This embodiment is advantageous in that the method relies on the detection of 15 the specific antibodies that are neutralizing for cell infection, that is those patient's antibodies that are effective against viraemia.

In a further embodiment of this invention, commercial diagnostic kits may be useful to carry out the above diagnosis methods, by detecting the presence or 20 absence of immune complexes formed by pestivirus particles and antibodies directed against pestivirus in a biological sample from a subject susceptible to be infected with pestivirus, or by detecting an inhibition of pestivirus-like particle infection of a permissive cell by anti-pestivirus neutralizing antibodies likely to be present in a biological sample of the patient. Such kits may comprise at least a pestivirus-like particle of the present invention. Where the method involves detection of immune 25 complexes, the kits may further comprise appropriate means of detection of said immune complexes. Preferably the kit of the invention further comprises directions, and protocols, depending upon the method selected, e.g., "competitive", "sandwich", and the like. The kits may also contain peripheral reagents such as buffers, stabilizers, etc...

30

In another aspect of the invention, the infectious pestivirus-like particles may be used for vaccination purposes.

According to an embodiment, the invention thus proposes a method of vaccination, notably against pestivirus infection, that comprises administration of a

pestivirus-like particle to a subject in need thereof. The invention also relates to a vaccine composition comprising a pestivirus-like particle and a pharmaceutically acceptable carrier. The invention further provides an immunogenic composition comprising in a pharmaceutical acceptable carrier, a pestivirus-like particle disclosed  
5 herein.

The vaccine and immunogenic compositions of the invention may be drawn to confer immunity, or elicit an immune response against pestivirus.

However, where the pestivirus-like particles of the invention further carry an additional gene encoding another antigen, different from pestivirus antigens, the  
10 invention provides a recombinant viral vaccine useful to raise an immune response against said antigen. Actually, the use of pseudo-particles described herein makes it possible to improve the elicited immune response through combining several presentation and processing pathways of an antigen. For instance, a vaccine composition of the invention, when administered, results in the pestivirus-like  
15 particles infecting cells of the host. The transgene encoding the antigen is then integrated in the cellular genome, and subsequently expressed by the cell, such that there is both a cellular and a humoral immune response elicited by the vaccine composition.

Advantageously, the pestivirus-like particles may further carry a transgene  
20 encoding an immune modulator, which allows for enhancement of the raised immune reaction.

The vaccination or immunogenic composition of the present invention may additionally contain an adjuvant. A number of adjuvants are known to those skilled in the art. Examples of suitable adjuvants include, for example, include aluminum  
25 hydroxide; Saponin; detergents such as Tween 80; animal, mineral or vegetable oils, *Corynebacterium* or *Propionibacterium* -derived adjuvants; *Mycobacterium bovis* (*Bacillus Calmette* and *Guerinn*, or BCG); cytokines; acrylic acid polymers such as carbomer; EMA; or combinations thereof.

The route of administration is any conventional route used in the vaccine field.  
30 As general guidance, a vaccine composition of the invention is administered via a mucosal surface, e.g., an ocular, intranasal, pulmonary, oral, intestinal, rectal, vaginal, and urinary tract surface; or via a parenteral route, e.g., by an intravenous, subcutaneous, intraperitoneal, intradermal, intraepidermal, or intramuscular route. The choice of administration route depends on the formulation that is selected.

In still another embodiment the particles of the invention may be used as vectors for gene transfer and/or gene therapy. Gene therapy is defined as the introduction of genetic material into a cell in order to either change its phenotype or genotype. Furthermore, such a delivery system is amenable to scale up for 5 reproducibly producing large titers of infectious, replication-defective pestivirus-like particles particles.

Accordingly, the invention relates to a method for *in vivo* or *in vitro* transferring a transgene of interest in a cell, which method comprises infecting a cell with a 10 pestivirus-like particle of the invention, wherein the particle carries a transgene of interest.

The invention further relates to the use of a pestivirus-like particle of the invention, that carries a transgene of interest, for the preparation of a medicament for the prevention or treatment of a disease in a patient, wherein the pestivirus-like 15 particle allows the transfer of the transgene of interest into a cell of the patient, and encodes a product that has a prophylactic or therapeutic effect against the disease.

In the above described uses of the particles of the invention the pestivirus may 20 preferably be selected from the group consisting of bovine viral diarrhea virus (BVDV), swine fever virus (SFV), and Border disease virus (BDV).

The invention will be further understood in view of the following examples.

**EXAMPLE 1 : Generation of pestivirus pseudo-particles**

Pestivirus pseudo-particles are generated by assembling full-length, 25 unmodified Erns, E1 and E2 glycoproteins onto retroviral core proteins derived from murine leukemia virus (MLV). To investigate further whether functional pestivirus pseudo-particles could also be produced with Erns, E1 and E2 expressed in *trans* with only one or two of the glycoproteins, expression vectors that encode Erns, E1, 30 E2, Erns and E1, Erns and E2, or Erns and E1 and E2 glycoproteins are designed.

*Construction of expression vectors encoding the viral components*

Plasmids expressing wild type ErnsE1E2 polyproteins are constructed by standard methods (Sambrook et al., 1989).

The specific polynucleotide and polypeptide constructs of BVDV deltaCErnsE1E2p7, deltaCErbsE1E2, deltaCE1E2p7 and deltaCE1E2 are shown in SEQ ID No19 to 26, respectively.

5 *Generation of pestivirus pseudo-particles*

Retroviruses were chosen as platforms for assembly of pestivirus-pp because their cores can incorporate a variety of different cellular and viral glycoproteins and because they can easily package and integrate genetic markers into host cell DNA.

10

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CLAIMS

1. A method for producing pestivirus-like particles *ex vivo* comprising the  
5 steps of:

- providing a first nucleic acid sequence comprising a packaging competent  
retroviral-derived genome;

- providing a second nucleic acid sequence comprising a cDNA encoding core  
proteins from said retrovirus;

10 - providing a third nucleic acid sequence comprising a cDNA encoding a  
polyprotein comprising successively a pestivirus core protein, and a Erns protein  
and/or a pestivirus E1 protein and/or a pestivirus E2 protein, and optionally a  
pestivirus p7 protein;

15 - transfecting host cells with said nucleic acid sequences and maintaining the  
transfected cells in culture for sufficient time to allow expression of the cDNAs to  
produce structural proteins from pestivirus and retrovirus; and allowing the structural  
proteins to form virus-like particles.

20 2. The method according to claim 1, wherein said packaging competent  
retroviral-derived genome and core proteins are from a retrovirus selected from the  
group consisting of MLV, ALV, RSV, MPMV, HIV-1, HIV-2, SIV, EIAV, CAEV, or  
HFV.

25 3. The method according to claim 1 or 2, wherein core, Erns, E1 and E2  
pestivirus proteins, and optionally p7 pestivirus protein, are derived from a same  
pestivirus.

4. The method according to any of preceding claims, wherein said  
pestivirus is selected from the group consisting of bovine viral diarrhea virus, swine  
fever virus, and border disease virus.

30 5. An infectious pestivirus-like particle susceptible to be obtained by a  
method according to any of preceding claims, comprising the core proteins from a  
retrovirus, and a Erns pestivirus protein and/or a E1 pestivirus protein and/or a E2  
pestivirus protein, and optionally a p7 pestivirus protein.

6. The infectious particle according to claim 5, wherein Erns, E1 protein  
and E2 protein, and optionally p7 pestivirus protein, are derived from a same  
pestivirus.

7. The infectious particle according to claim 6, wherein said pestivirus is selected from the group consisting of bovine viral diarrhea virus, swine fever virus, and border disease virus.

8. The infectious particle according to any of claims 5 to 7, wherein said retrovirus is selected from the group consisting of MLV, ALV, RSV, MPMV, HIV-1, HIV-2, SIV, EIAV, CAEV, or HFV.

9. Use of three nucleic acid sequences for the preparation of a medicament useful as a vaccine against a pestivirus infection, wherein the nucleic acid sequences are :

10 - a first nucleic acid sequence comprising a packaging competent retroviral-derived genome;

- a second nucleic acid sequence comprising a cDNA encoding core proteins from said retrovirus;

15 - a third nucleic acid sequence comprising a cDNA encoding a polyprotein comprising successively a pestivirus core protein, and a pestivirus Erns protein and/or a pestivirus E1 protein and/or a pestivirus E2 protein, and optionally a pestivirus p7 protein ;

20 and, when transferred into cells of a subject, the nucleic acids sequences allow the production of structural proteins from pestivirus and retrovirus, wherein the structural proteins form virus-like particles that are immunogenic.

10. The use according to claim 9, wherein said pestivirus is selected from the group consisting of bovine viral diarrhea virus, swine fever virus, and border disease virus.

BET 03/P0182

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**Abstract**

The invention relates to the generation and the use of pestivirus pseudo-particles containing native functional E1, E2 envelope glycoproteins assembled onto retroviral  
15 core particles. These particles are highly infectious and constitute a valid model of pestivirus virion.

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Val Lys Tyr Gln Val Arg Lys Lys Gly Lys Thr Lys Ser Lys Asn Thr  
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Glu Asp Gly Lys Leu Met Tyr Leu Gln Arg Cys Thr Arg Glu Thr Arg  
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Tyr Leu Ala Ile Leu His Thr Arg Ala Leu Pro Thr Ser Val Val Phe  
 65 70 75 80

Lys Lys Leu Phe Asp Gly Arg Lys Gln Glu Asp Val Val Glu Met Asn  
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Asp Asn Phe Glu Phe Gly Leu Cys Pro Cys Asp Ala Lys Pro Ile Val  
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Arg Gly Lys Phe Asn Thr Thr Leu Leu Asn Gly Pro Ala Phe Gln Met  
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Val Cys Pro Ile Gly Trp Thr Gly Thr Val Ser Cys Thr Ser Phe Asn  
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Pro Phe Pro His Arg Gln Gly Cys Ile Thr Gln Lys Asn Leu Gly Glu  
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Asp Leu His Asn Cys Ile Leu Gly Gly Asn Trp Thr Cys Val Pro Gly  
 180 185 190

Asp Gln Leu Leu Tyr Lys Gly Gly Ser Ile Glu Ser Cys Lys Trp Cys  
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Gly Tyr Gln Phe Lys Glu Ser Glu Gly Leu Pro His Tyr Pro Ile Gly  
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Arg Thr Lys Pro Pro Asp Ala Thr Ile Val Val Glu Gly Val Lys Tyr  
35 40 45

Gln Val Lys Lys Gly Lys Val Lys Gly Lys Ser Thr Gln Asp Gly  
50 55 60

Leu Tyr His Asn Lys Asn Lys Pro Pro Glu Ser Arg Lys Lys Leu Glu  
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Trp Pro Glu Lys Ile Cys Lys Gly Val Pro Thr Tyr Leu Ala Thr Asp  
35 40 45

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Thr Cys Arg Tyr Asp Lys Asp Ala Asp Ile Asn Val Val Thr Gln Ala  
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Val Glu Asp Ile Leu Tyr Gly Asp His Glu Cys Gly Ser Leu Leu Gln  
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Asp Thr Ala Leu Tyr Leu Val Asp Gly Met Thr Asn Thr Ile Glu Asn  
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Ala Arg Gln Gly Ala Ala Arg Val Thr Ser Trp Leu Gly Arg Gln Leu  
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Gly Pro Gly Lys Phe Asp Thr Asn Ala Glu Asp Gly Lys Ile Leu His

35

40

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Glu Met Gly Gly His Leu Ser Glu Phe Leu Leu Leu Ser Leu Val Val  
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Leu Ser Asp Phe Ala Pro Glu Thr Ala Ser Ala Leu Tyr Leu Ile Leu  
 65 70 75 80

His Tyr Met Ile Pro Gln Ser His Glu Glu Pro Glu Gly Cys Asp Thr  
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Asn Gln Leu Asn Leu Thr Val Glu Leu Arg Thr Glu Asp Val Ile Pro  
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Ser Ser Val Trp Asn Val Gly Lys Tyr Val Cys Val Arg Pro Asp Trp  
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Trp Pro Tyr Glu Thr Lys Val Ala Leu Leu Phe Glu Glu Ala Gly Gln  
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Val Val Lys Leu Ala Leu Arg Ala Leu Arg Asp Leu Thr Arg Val Trp  
 145 150 155 160

Asn Ser Ala Ser Thr Thr Ala Phe Leu Ile Cys Leu Ile Lys Val Leu  
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Ala Gln Gly  
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 20 25 30

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 35 40 45

Val Ala Gly Ser Phe Lys Val Ile Ala Leu Asn Val Val Ser Arg Arg  
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Tyr Leu Ala Ser Leu His Lys Glu Ala Ser Leu Thr Ser Val Thr Phe  
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Glu Leu Leu Phe Asp Gly Thr Asn Pro Ser Thr Glu Glu Met Gly Asp  
 85 90 95

Asp Phe Gly Phe Gly Leu Cys Pro Phe Asp Thr Ser Pro Val Val Lys  
 100 105 110

Gly Lys Tyr Asn Thr Thr Leu Leu Asn Gly Ser Ala Phe Tyr Leu Val  
 115 120 125

Cys Pro Ile Gly Trp Thr Gly Val Ile Glu Cys Thr Ala Val Ser Pro  
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Thr Thr Leu Arg Thr Glu Val Val Lys Thr Phe Arg Arg Asp Lys Pro  
 145 150 155 160

Phe Pro His Arg Met Asp Cys Ala Thr Thr Val Glu Asn Gly Asp  
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Leu Phe Tyr Cys Lys Leu Gly Gly Asn Trp Thr Cys Val Lys Gly Glu  
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Pro Val Val Tyr Thr Gly Gly Leu Val Lys Gln Cys Arg Trp Cys Gly  
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Phe Asp Phe Asn Glu Pro Asp Gly Leu Pro His Tyr Pro Ile Gly Lys  
 210 215 220

Cys Ile Leu Val Asn Glu Thr Gly Tyr Arg Ile Val Asp Ser Thr Asp  
 225 230 235 240

Cys Asn Arg Asp Gly Val Val Ile Ser Thr Asp Gly Ser His Glu Cys  
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Val Arg Lys Thr Ser Cys Thr Phe Asn Tyr Ala Lys Thr Leu Lys Asn  
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Lys Tyr Tyr Glu Pro Arg Asp Ser Tyr Phe Gln Gln Tyr Met Leu Lys  
 305 310 315 320

Gly Glu Tyr Gln Tyr Trp Phe Asp Leu Asp Val Thr Asp Arg His Ser  
 325 330 335

Asp Tyr Phe Ala Glu Phe Val Val Leu Val Val Val Ala Leu Leu Gly  
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8931

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9291

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10131

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<210> 14  
<211> 100  
<212> PRT  
<213> Border disease virus

<400> 14

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Val Arg Arg Gly Ala Met Lys Ile Thr Pro Lys Glu Ser Glu Lys Asp  
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Ser Arg Ser Lys Pro Pro Asp Ala Thr Ile Val Val Glu Gly Ile Lys  
35 40 45

Tyr Gln Val Lys Lys Gly Lys Val Lys Gly Lys Asn Thr Gln Asp  
50 55 60

Gly Leu Tyr His Asn Lys Asn Lys Pro Pro Glu Ser Arg Lys Lys Leu  
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Glu Lys Ala Leu Leu Ala Trp Ala Ile Ile Ala Ile Phe Met Trp Glu  
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Pro Val Ala Pro  
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<210> 15  
<211> 227  
<212> PRT  
<213> Border disease virus

<400> 15

Glu Asn Val Thr Gln Trp Asn Leu Ser Asp Asn Gly Thr Thr Gly Ile  
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Gln Leu Leu Met Phe Gln Arg Gly Val Asn Arg Ser Leu His Gly Ile  
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Trp Pro Glu Lys Ile Cys Thr Gly Val Pro Thr His Leu Ala Thr Asp  
 35 40 45

Ala Glu Leu Lys Gly Ile Gln Gly Met Met Asp Ala Ser Glu Lys Thr  
 50 55 60

Asn Tyr Thr Cys Cys Arg Leu Gln Arg His Glu Trp Asn Lys Tyr Gly  
 65 70 75 80

Trp Cys Asn Trp Tyr Asn Ile Asn Pro Trp Ile Trp Leu Met Asn Lys  
 85 90 95

Thr Gln Ala Asn Leu Thr Glu Gly Pro Pro Glu Lys Glu Cys Ala Val  
 100 105 110

Thr Cys Arg Phe Asp Lys Glu Ala Asp Ile Asn Ile Val Thr Gln Ala  
 115 120 125

Arg Asp Arg Pro Thr Thr Leu Thr Gly Cys Lys Lys Gly Lys Lys Phe  
 130 135 140

Ser Phe Ala Gly Met Ile Ile Glu Gly Pro Cys Asn Phe Asn Val Ser  
 145 150 155 160

Val Glu Asp Ile Leu Phe Gly Asp Asn Glu Cys Ser Ser Leu Phe Gln  
 165 170 175

Asp Thr Ala Leu Tyr Val Val Asp Gly Val Thr Asn Thr Val Glu Asn  
 180 185 190

Ala Arg Gln Gly Ala Ala Lys Leu Thr Ser Trp Leu Gly Lys Gln Leu  
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Gly Ile Met Gly Lys Lys Leu Glu His Lys Ser Lys Thr Trp Phe Gly  
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Ala Asn Ala  
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<210> 16  
 <211> 195  
 <212> PRT  
 <213> Border disease virus

<400> 16

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Gly Pro Gly Lys Phe Asp Thr Asn Ala Glu Asp Gly Lys Ile Leu His  
 35 40 45

Glu Met Arg Gly His Ile Ser Glu Phe Ile Leu Leu Ser Leu Val Val  
 50 55 60

Leu Ser Asp Phe Ala Pro Glu Thr Ala Ser Thr Leu Tyr Leu Val Leu  
 65 70 75 80

His Phe Ala Leu Pro Gln Thr His Glu Val Pro Ser Val Cys Asp Thr  
 85 90 95

Asn Gln Leu Asn Leu Thr Val Ser Leu Arg Val Asp Asp Val Ile Pro  
 100 105 110

Ser Ser Val Trp Asn Leu Gly Lys Tyr Val Cys Val Arg Pro Asp Trp  
 115 120 125

Trp Pro Tyr Glu Thr Thr Met Val Leu Leu Phe Glu Glu Ala Gly Gln  
 130 135 140

Val Val Lys Leu Val Leu Arg Ala Ile Arg Asp Leu Thr Arg Val Trp  
 145 150 155 160

Asn Ser Ala Ser Thr Thr Ala Phe Leu Ile Cys Leu Val Lys Val Leu  
 165 170 175

Arg Gly Gln Val Val Gln Gly Leu Val Trp Leu Leu Leu Val Thr Gly  
 180 185 190

Ala Gln Gly  
 195

<210> 17  
 <211> 373  
 <212> PRT  
 <213> Border disease virus

<400> 17

Gln Phe Ala Cys Arg Glu Asp Tyr Arg Tyr Ala Leu Ala Arg Thr Lys  
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Glu Ile Gly Ala Leu Gly Ala Glu Ser Leu Thr Thr Thr Trp Thr Asp  
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Tyr Arg Gly Asn Leu Glu Leu Asp Asp Gly Thr Val Arg Ala Thr Cys  
 35 40 45

Ser Arg Gly Phe Phe Arg Phe Arg Gly His Cys Met Ile Gly Pro Arg  
 50 55 60

Tyr Leu Ala Ser Leu His Leu Arg Ala Leu Pro Thr Ser Val Thr Phe  
 65 70 75 80

Glu Leu Ile Pro Gly Gly Ser Ala Met Thr Glu Glu Glu Met Gly Asp  
 85 90 95

Asp Phe Glu Phe Gly Leu Cys Pro Cys Asp Ser Arg Pro Val Val Lys  
 100 105 110

Gly Lys Tyr Asn Thr Thr Leu Leu Asn Gly Ser Ala Phe Gln Leu Ile  
 115 120 125

Cys Pro Tyr Gly Trp Val Gly Arg Val Glu Cys Thr Thr Val Ser Lys  
 130 135 140

Ser Thr Leu Ala Thr Glu Val Val Lys Ile Tyr Lys Lys Thr Lys Pro  
 145 150 155 160

Phe Pro Gln Arg Val Gly Cys Asp His Thr Thr Val Tyr Lys Gln Asp  
 165 170 175

Leu Tyr His Cys Gln Met Gly Gly Asn Trp Thr Cys Met Arg Gly Glu  
 180 185 190

Val Val Lys Tyr Val Gly Gly Pro Val Lys Lys Cys Glu Trp Cys Gly  
 195 200 205

Tyr Val Phe Lys Lys Arg Glu Gly Leu Pro His Tyr Pro Ile Gly Arg  
 210 215 220

Cys Met Leu Arg Asn Glu Thr Gly Tyr Arg Ser Val Asp Asp Thr Pro  
 225 230 235 240

Cys Asp Arg Gly Gly Val Val Ile Ser Lys Thr Gly Glu Leu Glu Cys  
 245 250 255

Leu Ile Gly Lys Thr Thr Val Lys Val Phe Ser Ser Asp Lys Lys Leu  
 260 265 270

Gly Pro Met Pro Cys Arg Pro Lys Glu Val Ile Ser Ser Glu Gly Pro  
 275 280 285

Val Ser Lys Ile Ala Cys Thr Phe Asn Tyr Ser Lys Thr Leu Glu Asn  
 290 295 300

Lys Tyr Tyr Glu Pro Arg Asp Ser Tyr Phe Gln Gln Tyr Met Leu Lys  
 305 310 315 320

Gly Gln Tyr Gln Tyr Trp Phe Asp Leu Glu Ala Thr Asp His His Ser  
 325 330 335

Asp Tyr Phe Ala Glu Phe Ile Met Leu Ala Val Val Ala Leu Leu Gly  
 340 345 350

Gly Arg Tyr Val Leu Trp Leu Met Val Val Tyr Met Ile Leu Ala Asp  
 355 360 365

Gln Met Thr Ser Ala  
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<210> 18  
 <211> 70  
 <212> PRT  
 <213> Border disease virus

<400> 18

Ile Asn Leu Gly Gln Gly Glu Val Val Leu Ile Gly Asn Leu Ile Thr  
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His Glu Asp His Glu Val Val Tyr Phe Leu Leu Leu Tyr Leu Ile  
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Val Lys Asp Glu Pro Val Lys Lys Trp Ile Leu Phe Leu Phe His Ala  
 35 40 45

Met Thr Asn Asn Pro Val Lys Thr Ile Ser Val Gly Leu Leu Met Leu  
 50 55 60

Ser Gly Leu Val Lys Gly  
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<210> 19

<211> 2694  
<212> DNA  
<213> Bovine viral diarrhea virus : deltaCErnsE1E2p7

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120

gataatggg cggaaaggat acaacggca atgttccaa ggggtgtgaa tagaagttt  
180

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240

gaactaaaaa caattcatgg tatgtatggat gcaagtgaga agaccaacta cacgtgtgc  
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360

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420

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540

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660

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720

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900

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960

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1080

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<210> 20
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<212> PRT
<213> Bovine viral diarrhea virus : deltaCErnsE1E2p7
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<223> E1
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Met Gly Glu Asn Ile Thr Gln Trp Asn Leu Gln Asp Asn Gly Thr Glu  
 35 40 45

Gly Ile Gln Arg Ala Met Phe Gln Arg Gly Val Asn Arg Ser Leu His  
 50 55 60

Gly Ile Trp Pro Glu Lys Ile Cys Thr Gly Val Pro Ser His Leu Ala  
 65 70 75 80

Thr Asp Ile Glu Leu Lys Thr Ile His Gly Met Met Asp Ala Ser Glu  
 85 90 95

Lys Thr Asn Tyr Thr Cys Cys Arg Leu Gln Arg His Glu Trp Asn Lys  
 100 105 110

His Gly Trp Cys Asn Trp Tyr Asn Ile Glu Pro Trp Ile Leu Val Met  
 115 120 125

Asn Arg Thr Gln Ala Asn Leu Thr Glu Gly Gln Pro Pro Arg Glu Cys  
 130 135 140

Ala Val Thr Cys Arg Tyr Asp Arg Ala Ser Asp Leu Asn Val Val Thr  
 145 150 155 160

Gln Ala Arg Asp Ser Pro Thr Pro Leu Thr Gly Cys Lys Lys Gly Lys  
 165 170 175

Asn Phe Ser Phe Ala Gly Ile Leu Met Arg Gly Pro Cys Asn Phe Glu  
 180 185 190

Ile Ala Ala Ser Asp Val Leu Phe Lys Glu His Glu Arg Ile Ser Met  
 195 200 205

Phe Gln Asp Thr Thr Leu Tyr Leu Val Asp Gly Leu Thr Asn Ser Leu  
 210 215 220

Glu Gly Ala Arg Gln Gly Thr Ala Lys Leu Thr Thr Trp Leu Gly Lys  
 225 230 235 240

Gln Leu Gly Ile Leu Gly Lys Lys Leu Glu Asn Lys Ser Lys Thr Trp  
 245 250 255

Phe Gly Ala Tyr Ala Ala Ser Pro Tyr Cys Asp Val Asp Arg Lys Ile

260

265

270

Gly Tyr Ile Trp Tyr Thr Lys Asn Cys Thr Pro Ala Cys Leu Pro Lys  
 275 280 285

Asn Thr Lys Ile Val Gly Pro Gly Lys Phe Gly Thr Asn Ala Glu Asp  
290 295 300

Gly Lys Ile Leu His Glu Met Gly Gly His Leu Ser Glu Val Leu Leu  
 305 310 315 320

Leu Ser Leu Val Val Leu Ser Asp Phe Ala Pro Glu Thr Ala Ser Val  
325 330 335

Met Tyr Leu Ile Leu His Phe Ser Ile Pro Gln Ser His Val Asp Val  
340 345 350

Met Asp Cys Asp Lys Thr Gln Leu Asn Leu Thr Val Glu Leu Thr Thr  
 355 360 365

Ala Glu Val Ile Pro Gly Ser Val Trp Asn Leu Gly Lys Tyr Val Cys  
370 375 380

Ile Arg Pro Asn Trp Trp Pro Tyr Glu Thr Thr Val Val Leu Ala Phe  
 385 390 395 400

Glu Glu Val Ser Gln Val Val Lys Leu Val Leu Arg Ala Leu Arg Asp  
 405 410 415

Leu Thr Arg Ile Trp Asn Ala Ala Thr Thr Thr Ala Phe Leu Val Cys  
420 425 430

Leu Val Lys Ile Val Arg Gly Gln Met Val Gln Gly Ile Leu Trp Leu  
435 440 445

Leu Leu Ile Thr Gly Val Gln Gly His Leu Asp Cys Lys Pro Glu Phe  
450 455 460

Ser Tyr Ala Ile Ala Lys Asp Glu Arg Ile Gly Gln Leu Gly Ala Glu  
 465 470 475 480

Gly Leu Thr Thr Trp Lys Glu Tyr Ser Pro Gly Met Lys Leu Glu  
 485 490 495

Asp Thr Met Val Ile Ala Trp Cys Glu Asp Gly Lys Leu Met Tyr Leu  
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Gln Arg Cys Thr Arg Glu Thr Arg Tyr Leu Ala Ile Leu His Thr Arg  
 515 520 525

Ala Leu Pro Thr Ser Val Val Phe Lys Lys Leu Phe Asp Gly Arg Lys  
 530 535 540

Gln Glu Asp Val Val Glu Met Asn Asp Asn Phe Glu Phe Gly Leu Cys  
 545 550 555 560

Pro Cys Asp Ala Lys Pro Ile Val Arg Gly Lys Phe Asn Thr Thr Leu  
 565 570 575

Leu Asn Gly Pro Ala Phe Gln Met Val Cys Pro Ile Gly Trp Thr Gly  
 580 585 590

Thr Val Ser Cys Thr Ser Phe Asn Met Asp Thr Leu Ala Thr Thr Val  
 595 600 605

Val Arg Thr Tyr Arg Arg Ser Lys Pro Phe Pro His Arg Gln Gly Cys  
 610 615 620

Ile Thr Gln Lys Asn Leu Gly Glu Asp Leu His Asn Cys Ile Leu Gly  
 625 630 635 640

Gly Asn Trp Thr Cys Val Pro Gly Asp Gln Leu Leu Tyr Lys Gly Gly  
 645 650 655

Ser Ile Glu Ser Cys Lys Trp Cys Gly Tyr Gln Phe Lys Glu Ser Glu  
 660 665 670

Gly Leu Pro His Tyr Pro Ile Gly Lys Cys Lys Leu Glu Asn Glu Thr  
 675 680 685

Gly Tyr Arg Leu Val Asp Ser Thr Ser Cys Asn Arg Glu Gly Val Ala  
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Ile Val Pro Gln Gly Thr Leu Lys Cys Lys Ile Gly Lys Thr Thr Val  
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Gln Val Ile Ala Met Asp Thr Lys Leu Gly Pro Met Pro Cys Arg Pro  
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Tyr Glu Ile Ile Ser Ser Glu Gly Pro Val Glu Lys Thr Ala Cys Thr  
 740 745 750

Phe Asn Tyr Thr Lys Thr Leu Lys Asn Lys Tyr Phe Glu Pro Arg Asp  
 755 760 765

Ser Tyr Phe Gln Gln Tyr Met Leu Lys Gly Glu Tyr Gln Tyr Trp Phe  
 770 775 780

Asp Leu Glu Val Thr Asp His His Arg Asp Tyr Phe Ala Glu Ser Ile  
 785 790 795 800

Leu Val Val Val Ala Leu Leu Gly Gly Arg Tyr Val Leu Trp Leu  
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Leu Val Thr Tyr Met Val Leu Ser Glu Gln Lys Ala Leu Gly Ile Gln  
 820 825 830

Tyr Gly Ser Gly Glu Val Val Met Met Gly Asn Leu Leu Thr His Asn  
 835 840 845

Asn Ile Glu Val Val Thr Tyr Phe Leu Leu Leu Tyr Leu Leu Leu Arg  
 850 855 860

Glu Glu Ser Val Lys Lys Trp Val Leu Leu Leu Tyr His Ile Leu Val  
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Val Val Lys Ala  
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<210> 21

<211> 2484

<212> DNA

<213> Bovine viral diarrhea virus : delraCErnsE1E2

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 240

gaactaaaaa caattcatgg tatgatggat gcaagtgaga agacccaacta cacgtgtgc  
 300

agacttcaac gccatgagtg gaacaagcat ggttggtgca actggcacaa tattgaaccc  
360

tggattctag tcatgaatag aacccaagcc aatctcaactg agggacaacc accaaggag  
420

tgcgcagtca cttgttaggta tgataggct agtacttaa acgtggtaac acaagctaga  
480

gatagccccca cacccttaac aggttgcaag aaaggaaaga acttctcctt tgcaggcata  
540

ttgatgcggg gcccctgcaa ctttgaataa gctgcaagtg atgtattatt caaagaacat  
600

gaacgcatta gtatgttcca ggataccact cttaaccttg ttgacggggtt gaccaactcc  
660

ttagaagggtg ccagacaagg aaccgctaaa ctgacaacct ggttaggcaaa gcagctcggg  
720

atactaggaa aaaagttgga aaacaagagt aagacgtggt ttggagcata cgctgcttcc  
780

ccttactgtg atgtcgatcg caaaattggc tacatatggt atacaaaaaaa ttgcacccct  
840

gcctgcttac ccaagaacac aaaaattgtc ggccctggaa aatttggcac caatgcagag  
900

gacggcaaga tattacatga gatgggggtt cacttgcgg aggtactact actttcttta  
960

gtggtgctgt ccgacttcgc accggaaaca gctagtgtaa tgtacctaattt cctacatttt  
1020

tccatcccac aaagtcacgt tgatgtaatg gattgtgata agacccagtt gaacctcaca  
1080

gtggagctga caacagctga agtaataccaa gggtcggctt ggaatctagg caaatatgt  
1140

tgtataagac caaattggtg gccttatgag acaactgttag tggggcatt tgaagaggt  
1200

agccaggtgg tgaagtttgtt gttgagggca ctcagagatt taacacgcatt ttggAACGCT  
1260

gcaacaacta ctgtttttt agtatgcctt gttaagatag tcagggccaa gatggtag  
1320

ggcattctgt ggctactatt gataacaggg gtacaaggc acttggattt gaaacctgaa  
1380

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1440

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 1680  
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 1740  
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 1800  
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 1920  
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 1980  
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 2100  
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 2160  
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 2280  
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 2340  
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 2400  
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<210> 22  
 <211> 827  
 <212> PRT  
 <213> Bovine viral diarrhea virus : deltaCErnsE1E2  
 <400> 22

Lys Asn Lys Pro Gln Glu Ser Arg Lys Lys Leu Glu Lys Ala Leu Leu  
 1 5 10 15

Ala Trp Ala Ile Ile Ala Ile Val Leu Phe Gln Val Thr Met Gly Glu  
20 25 30

Asn Ile Thr Gln Trp Asn Leu Gln Asp Asn Gly Thr Glu Gly Ile Gln  
35 40 45

Arg Ala Met Phe Gln Arg Gly Val Asn Arg Ser Leu His Gly Ile Trp  
50 55 60

Pro Glu Lys Ile Cys Thr Gly Val Pro Ser His Leu Ala Thr Asp Ile  
65 70 75 80

Glu Leu Lys Thr Ile His Gly Met Met Asp Ala Ser Glu Lys Thr Asn  
85 90 95

Tyr Thr Cys Cys Arg Leu Gln Arg His Glu Trp Asn Lys His Gly Trp  
100 105 110

Cys Asn Trp Tyr Asn Ile Glu Pro Trp Ile Leu Val Met Asn Arg Thr  
115 120 125

Gln Ala Asn Leu Thr Glu Gly Gln Pro Pro Arg Glu Cys Ala Val Thr  
130 135 140

Cys Arg Tyr Asp Arg Ala Ser Asp Leu Asn Val Val Thr Gln Ala Arg  
145 150 155 160

Asp Ser Pro Thr Pro Leu Thr Gly Cys Lys Lys Gly Lys Asn Phe Ser  
165 170 175

Phe Ala Gly Ile Leu Met Arg Gly Pro Cys Asn Phe Glu Ile Ala Ala  
180 185 190

Ser Asp Val Leu Phe Lys Glu His Glu Arg Ile Ser Met Phe Gln Asp  
195 200 205

Thr Thr Leu Tyr Leu Val Asp Gly Leu Thr Asn Ser Leu Glu Gly Ala  
210 215 220

Arg Gln Gly Thr Ala Lys Leu Thr Thr Trp Leu Gly Lys Gln Leu Gly  
225 230 235 240

Ile Leu Gly Lys Lys Leu Glu Asn Lys Ser Lys Thr Trp Phe Gly Ala  
245 250 255

Tyr Ala Ala Ser Pro Tyr Cys Asp Val Asp Arg Lys Ile Gly Tyr Ile  
 260 265 270

Trp Tyr Thr Lys Asn Cys Thr Pro Ala Cys Leu Pro Lys Asn Thr Lys  
 275 280 285

Ile Val Gly Pro Gly Lys Phe Gly Thr Asn Ala Glu Asp Gly Lys Ile  
 290 295 300

Leu His Glu Met Gly Gly His Leu Ser Glu Val Leu Leu Leu Ser Leu  
 305 310 315 320

Val Val Leu Ser Asp Phe Ala Pro Glu Thr Ala Ser Val Met Tyr Leu  
 325 330 335

Ile Leu His Phe Ser Ile Pro Gln Ser His Val Asp Val Met Asp Cys  
 340 345 350

Asp Lys Thr Gln Leu Asn Leu Thr Val Glu Leu Thr Thr Ala Glu Val  
 355 360 365

Ile Pro Gly Ser Val Trp Asn Leu Gly Lys Tyr Val Cys Ile Arg Pro  
 370 375 380

Asn Trp Trp Pro Tyr Glu Thr Thr Val Val Leu Ala Phe Glu Glu Val  
 385 390 395 400

Ser Gln Val Val Lys Leu Val Leu Arg Ala Leu Arg Asp Leu Thr Arg  
 405 410 415

Ile Trp Asn Ala Ala Thr Thr Ala Phe Leu Val Cys Leu Val Lys  
 420 425 430

Ile Val Arg Gly Gln Met Val Gln Gly Ile Leu Trp Leu Leu Leu Ile  
 435 440 445

Thr Gly Val Gln Gly His Leu Asp Cys Lys Pro Glu Phe Ser Tyr Ala  
 450 455 460

Ile Ala Lys Asp Glu Arg Ile Gly Gln Leu Gly Ala Glu Gly Leu Thr  
 465 470 475 480

Thr Thr Trp Lys Glu Tyr Ser Pro Gly Met Lys Leu Glu Asp Thr Met  
 485 490 495

Val Ile Ala Trp Cys Glu Asp Gly Lys Leu Met Tyr Leu Gln Arg Cys  
 500 505 510

Thr Arg Glu Thr Arg Tyr Leu Ala Ile Leu His Thr Arg Ala Leu Pro  
 515 520 525

Thr Ser Val Val Phe Lys Lys Leu Phe Asp Gly Arg Lys Gln Glu Asp  
 530 535 540

Val Val Glu Met Asn Asp Asn Phe Glu Phe Gly Leu Cys Pro Cys Asp  
 545 550 555 560

Ala Lys Pro Ile Val Arg Gly Lys Phe Asn Thr Thr Leu Leu Asn Gly  
 565 570 575

Pro Ala Phe Gln Met Val Cys Pro Ile Gly Trp Thr Gly Thr Val Ser  
 580 585 590

Cys Thr Ser Phe Asn Met Asp Thr Leu Ala Thr Thr Val Val Arg Thr  
 595 600 605

Tyr Arg Arg Ser Lys Pro Phe Pro His Arg Gln Gly Cys Ile Thr Gln  
 610 615 620

Lys Asn Leu Gly Glu Asp Leu His Asn Cys Ile Leu Gly Gly Asn Trp  
 625 630 635 640

Thr Cys Val Pro Gly Asp Gln Leu Leu Tyr Lys Gly Gly Ser Ile Glu  
 645 650 655

Ser Cys Lys Trp Cys Gly Tyr Gln Phe Lys Glu Ser Glu Gly Leu Pro  
 660 665 670

His Tyr Pro Ile Gly Lys Cys Lys Leu Glu Asn Glu Thr Gly Tyr Arg  
 675 680 685

Leu Val Asp Ser Thr Ser Cys Asn Arg Glu Gly Val Ala Ile Val Pro  
 690 695 700

Gln Gly Thr Leu Lys Cys Lys Ile Gly Lys Thr Thr Val Gln Val Ile  
 705 710 715 720

Ala Met Asp Thr Lys Leu Gly Pro Met Pro Cys Arg Pro Tyr Glu Ile  
 725 730 735

Ile Ser Ser Glu Gly Pro Val Glu Lys Thr Ala Cys Thr Phe Asn Tyr

740	745	750
-----	-----	-----

Thr Lys Thr Leu Lys Asn Lys Tyr Phe Glu Pro Arg Asp Ser Tyr Phe  
 755 760 765

Gln Gln Tyr Met Leu Lys Gly Glu Tyr Gln Tyr Trp Phe Asp Leu Glu  
 770 775 780

Val Thr Asp His His Arg Asp Tyr Phe Ala Glu Ser Ile Leu Val Val  
 785 790 795 800

Val Val Ala Leu Leu Gly Gly Arg Tyr Val Leu Trp Leu Leu Val Thr  
 805 810 815

Tyr Met Val Leu Ser Glu Gln Lys Ala Leu Gly  
 820 825

<210> 23  
 <211> 2013  
 <212> DNA  
 <213> Bovine viral diarrhea virus : deltaCE1E2p7

<400> 23  
 aaaaaacaaac ctcaggaatc acgcaagaaa ctggaaaaag cattgttggc gtgggcaata  
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atagctatag ttttgttca agttacaatg ggagcttccc cttactgtga tgtcgatcgc  
 120

aaaattggct acatatggta tacaaaaaat tgcacccctg cctgcttacc caagaacaca  
 180

aaaattgtcg gccctggaa atttggcacc aatgcagagg acggcaagat attacatgag  
 240

atggggggtc acttgtcgga ggtactacta ctttctttag tggtgctgtc cgacttcgca  
 300

ccggaaacag ctatgtaat gtacctaattc ctacatttt ccatcccaca aagtcacgtt  
 360

gatgtaatgg attgtataa gaccctgtt aacctcacag tggagctgac aacagctgaa  
 420

gtaataccag ggtcggtctg gaatctaggc aaatatgtat gtataagacc aaattggtgg  
 480

ccttatgaga caactgttagt gttggcattt gaagaggtga gccaggtggt gaagtttagtg  
 540

ttgagggcac tcagagattt aacacgcatt tggAACGCTG caacaactac tgcttttta  
 600

gtatgccttg ttaagatagt caggggccag atggcacagg gcattctgtg gctactattg  
 660

ataaacagggg tacaaggcga cttggattgc aaacctgaat tctcgatgc catagcaaag  
720  
gacgaaagaa ttggtaact ggggctgaa ggccttacca ccacttgaa ggaatactca  
780  
cctggaatga agctggaaga cacaatggc attgcttggt gcgaagatgg gaagttaatg  
840  
tacctccaaa gatgcacgag agaaaccaga tatctcgcaa tcttcatac aagagcctg  
900  
ccgaccagtg tggattcaa aaaactctt gatggcgaa agcaagagga tggatcgaa  
960  
atgaacgaca actttgaatt tggactctgc ccatgtgatg ccaaaccat agtaagaggg  
1020  
aagttcaata caacgctgct gaacggaccg gccttccaga tggatgccc cataggatgg  
1080  
acagggactg taagctgtac gtcattcaat atggacacct tagccacaac tggatcgaa  
1140  
acatatacaa ggtctaaacc attccctcat aggcaaggct gtatcacca aaagaatctg  
1200  
ggggaggatc tccataactg catccttggaa ggaaatttggaa cttgtgtgcc tggagaccaa  
1260  
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1320  
agtggggac taccacacta ccccatggc aagtgtaaat tggagaacga gactggttac  
1380  
aggctagtag acagtaccc ttgcaataga gaagggttgg ccatagtacc acaagggaca  
1440  
ttaaagtgc aagatggaaa aacaactgta caggtcatag ctatggatac caaactcgga  
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cctatgcctt gcagaccata tggaaatcata tcaagtgggg ggcctgtaga aaagacagcg  
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1620  
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1680  
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1740  
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1800  
attcagttatg gatcaggggaa agtgggtatg atgggcaact tgctaaacca taacaatatt  
1860

gaagtggta catacttctt gctgctgtac ctactgctga gggaggagag cgtaaagaag  
1920

tgggtttac tcttatacca catcttagtg gtacacccaa tcaaatttgt aatttgtatc  
1980

ctactgatga ttggggatgt ggtaaaggcc tga  
2013

<210> 24

<211> 670

<212> PRT

<213> Bovine viral diarrhea virus : deltaCE1E2p7

<400> 24

Lys Asn Lys Pro Gln Glu Ser Arg Lys Lys Leu Glu Lys Ala Leu Leu  
1 5 10 15

Ala Trp Ala Ile Ile Ala Ile Val Leu Phe Gln Val Thr Met Gly Ala  
20 25 30

Ser Pro Tyr Cys Asp Val Asp Arg Lys Ile Gly Tyr Ile Trp Tyr Thr  
35 40 45

Lys Asn Cys Thr Pro Ala Cys Leu Pro Lys Asn Thr Lys Ile Val Gly  
50 55 60

Pro Gly Lys Phe Gly Thr Asn Ala Glu Asp Gly Lys Ile Leu His Glu  
65 70 75 80

Met Gly Gly His Leu Ser Glu Val Leu Leu Ser Leu Val Val Leu  
85 90 95

Ser Asp Phe Ala Pro Glu Thr Ala Ser Val Met Tyr Leu Ile Leu His  
100 105 110

Phe Ser Ile Pro Gln Ser His Val Asp Val Met Asp Cys Asp Lys Thr  
115 120 125

Gln Leu Asn Leu Thr Val Glu Leu Thr Thr Ala Glu Val Ile Pro Gly  
130 135 140

Ser Val Trp Asn Leu Gly Lys Tyr Val Cys Ile Arg Pro Asn Trp Trp  
145 150 155 160

Pro Tyr Glu Thr Thr Val Val Leu Ala Phe Glu Glu Val Ser Gln Val  
165 170 175

Val Lys Leu Val Leu Arg Ala Leu Arg Asp Leu Thr Arg Ile Trp Asn  
 180 185 190

Ala Ala Thr Thr Ala Phe Leu Val Cys Leu Val Lys Ile Val Arg  
 195 200 205

Gly Gln Met Val Gln Gly Ile Leu Trp Leu Leu Leu Ile Thr Gly Val  
 210 215 220

Gln Gly His Leu Asp Cys Lys Pro Glu Phe Ser Tyr Ala Ile Ala Lys  
 225 230 235 240

Asp Glu Arg Ile Gly Gln Leu Gly Ala Glu Gly Leu Thr Thr Trp  
 245 250 255

Lys Glu Tyr Ser Pro Gly Met Lys Leu Glu Asp Thr Met Val Ile Ala  
 260 265 270

Trp Cys Glu Asp Gly Lys Leu Met Tyr Leu Gln Arg Cys Thr Arg Glu  
 275 280 285

Thr Arg Tyr Leu Ala Ile Leu His Thr Arg Ala Leu Pro Thr Ser Val  
 290 295 300

Val Phe Lys Lys Leu Phe Asp Gly Arg Lys Gln Glu Asp Val Val Glu  
 305 310 315 320

Met Asn Asp Asn Phe Glu Phe Gly Leu Cys Pro Cys Asp Ala Lys Pro  
 325 330 335

Ile Val Arg Gly Lys Phe Asn Thr Thr Leu Leu Asn Gly Pro Ala Phe  
 340 345 350

Gln Met Val Cys Pro Ile Gly Trp Thr Gly Thr Val Ser Cys Thr Ser  
 355 360 365

Phe Asn Met Asp Thr Leu Ala Thr Thr Val Val Arg Thr Tyr Arg Arg  
 370 375 380

Ser Lys Pro Phe Pro His Arg Gln Gly Cys Ile Thr Gln Lys Asn Leu  
 385 390 395 400

Gly Glu Asp Leu His Asn Cys Ile Leu Gly Gly Asn Trp Thr Cys Val  
 405 410 415

Pro Gly Asp Gln Leu Leu Tyr Lys Gly Ser Ile Glu Ser Cys Lys

420	425	430
Trp Cys Gly Tyr Gln Phe Lys Glu Ser Glu Gly Leu Pro His Tyr Pro		
435	440	445
Ile Gly Lys Cys Lys Leu Glu Asn Glu Thr Gly Tyr Arg Leu Val Asp		
450	455	460
Ser Thr Ser Cys Asn Arg Glu Gly Val Ala Ile Val Pro Gln Gly Thr		
465	470	475
Leu Lys Cys Lys Ile Gly Lys Thr Thr Val Gln Val Ile Ala Met Asp		
485	490	495
Thr Lys Leu Gly Pro Met Pro Cys Arg Pro Tyr Glu Ile Ile Ser Ser		
500	505	510
Glu Gly Pro Val Glu Lys Thr Ala Cys Thr Phe Asn Tyr Thr Lys Thr		
515	520	525
Leu Lys Asn Lys Tyr Phe Glu Pro Arg Asp Ser Tyr Phe Gln Gln Tyr		
530	535	540
Met Leu Lys Gly Glu Tyr Gln Tyr Trp Phe Asp Leu Glu Val Thr Asp		
545	550	555
His His Arg Asp Tyr Phe Ala Glu Ser Ile Leu Val Val Val Ala		
565	570	575
Leu Leu Gly Gly Arg Tyr Val Leu Trp Leu Leu Val Thr Tyr Met Val		
580	585	590
Leu Ser Glu Gln Lys Ala Leu Gly Ile Gln Tyr Gly Ser Gly Glu Val		
595	600	605
Val Met Met Gly Asn Leu Leu Thr His Asn Asn Ile Glu Val Val Thr		
610	615	620
Tyr Phe Leu Leu Leu Tyr Leu Leu Leu Arg Glu Glu Ser Val Lys Lys		
625	630	635
Trp Val Leu Leu Leu Tyr His Ile Leu Val Val His Pro Ile Lys Ser		
645	650	655
Val Ile Val Ile Leu Leu Met Ile Gly Asp Val Val Lys Ala		
660	665	670

<210> 25  
<211> 1803  
<212> DNA  
<213> Bovine viral diarrhea virus : deltaCE1E2

<400> 25  
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120  
aaaattggct acatatggta tacaaaaaaat tgcacccctg cctgcttacc caagaacaca  
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240  
atggggggtc acttgcgga ggtactacta ctttcttag tggtgctgtc cgacttcgca  
300  
ccgaaacag ctatgtaat gtacctaatc ctacatccccccaca aagtacgtt  
360  
gatgtaatgg attgtgataa gaccagttg aacctcacag tggagctgac aacagctgaa  
420  
gtaataccag ggccggctcg gaatctaggc aaatatgtat gtataagacc aaattgggtgg  
480  
ccttatgaga caactgttagt gttggcattt gaagaggtga gccaggtggt gaagtttagtg  
540  
ttgagggcac tcagagattt aacacgcatt tggaaacgctg caacaactac tgctttttta  
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gtatgccttg ttaagatagt cagggccag atggcacagg gcattctgtg gctactattg  
660  
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720  
gacgaaagaa ttggtaact gggggctgaa ggccttacca ccactggaa ggaatactca  
780  
cctggaatga agctggaaaga cacaatggc attgcttggt gccaagatgg gaagttaatg  
840  
tacctccaaa gatgcacgag agaaaccaga tatctcgaa tcttcatac aagagccttg  
900  
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960  
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1020  
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1080

acagggactg taagctgtac gtcattcaat atggacacct tagccacaac tgtggtacgg  
1140

acatatagaa ggtctaaacc attccctcat aggcaaggct gtatcaccca aaagaatctg  
1200

ggggaggatc tccataactg catccttggaa ggaaatttggaa cttgtgtgcc tggagaccaa  
1260

ctactataca aagggggctc tattgaatct tgcaagtggt gtggctatca atttaaagag  
1320

agtgagggac taccacacta ccccatggc aagtgtaaat tggagaacga gactggttac  
1380

aggctagtag acagtacctc ttgcaataga gaagggtgtgg ccatagtacc acaagggaca  
1440

ttaaagtgc aagataggaaa aacaactgta caggtcatag ctatggatac caaactcgga  
1500

cctatgcctt gcagaccata taaaatcata tcaagtgagg ggcctgtaga aaagacagcg  
1560

tgtactttca actacactaa gacattaaaa aataagtatt ttgagccag agacagctac  
1620

tttcagcaat acatgctaaa aggagagtat caatactggt ttgacctgga ggtgactgac  
1680

catcaccggg attacttcgc tgagtccata ttagtggtgg tagtagccct cttgggtggc  
1740

agatatgtac tttggttact ggttacatac atggctttat cagaacagaa ggccttaggg  
1800

tga  
1803

<210> 26  
<211> 600  
<212> PRT  
<213> Bovine viral diarrhea virus : deltaCE1E2

<400> 26

Lys Asn Lys Pro Gln Glu Ser Arg Lys Lys Leu Glu Lys Ala Leu Leu  
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Ala Trp Ala Ile Ile Ala Ile Val Leu Phe Gln Val Thr Met Gly Ala  
20 25 30

Ser Pro Tyr Cys Asp Val Asp Arg Lys Ile Gly Tyr Ile Trp Tyr Thr  
35 40 45

Lys Asn Cys Thr Pro Ala Cys Leu Pro Lys Asn Thr Lys Ile Val Gly

50

55

60

Pro Gly Lys Phe Gly Thr Asn Ala Glu Asp Gly Lys Ile Leu His Glu  
 65 70 75 80

Met Gly Gly His Leu Ser Glu Val Leu Leu Ser Leu Val Val Leu  
 85 90 95

Ser Asp Phe Ala Pro Glu Thr Ala Ser Val Met Tyr Leu Ile Leu His  
 100 105 110

Phe Ser Ile Pro Gln Ser His Val Asp Val Met Asp Cys Asp Lys Thr  
 115 120 125

Gln Leu Asn Leu Thr Val Glu Leu Thr Thr Ala Glu Val Ile Pro Gly  
 130 135 140

Ser Val Trp Asn Leu Gly Lys Tyr Val Cys Ile Arg Pro Asn Trp Trp  
 145 150 155 160

Pro Tyr Glu Thr Thr Val Val Leu Ala Phe Glu Glu Val Ser Gln Val  
 165 170 175

Val Lys Leu Val Leu Arg Ala Leu Arg Asp Leu Thr Arg Ile Trp Asn  
 180 185 190

Ala Ala Thr Thr Ala Phe Leu Val Cys Leu Val Lys Ile Val Arg  
 195 200 205

Gly Gln Met Val Gln Gly Ile Leu Trp Leu Leu Ile Thr Gly Val  
 210 215 220

Gln Gly His Leu Asp Cys Lys Pro Glu Phe Ser Tyr Ala Ile Ala Lys  
 225 230 235 240

Asp Glu Arg Ile Gly Gln Leu Gly Ala Glu Gly Leu Thr Thr Trp  
 245 250 255

Lys Glu Tyr Ser Pro Gly Met Lys Leu Glu Asp Thr Met Val Ile Ala  
 260 265 270

Trp Cys Glu Asp Gly Lys Leu Met Tyr Leu Gln Arg Cys Thr Arg Glu  
 275 280 285

Thr Arg Tyr Leu Ala Ile Leu His Thr Arg Ala Leu Pro Thr Ser Val  
 290 295 300

Val Phe Lys Lys Leu Phe Asp Gly Arg Lys Gln Glu Asp Val Val Glu  
 305 310 315 320

Met Asn Asp Asn Phe Glu Phe Gly Leu Cys Pro Cys Asp Ala Lys Pro  
 325 330 335

Ile Val Arg Gly Lys Phe Asn Thr Thr Leu Leu Asn Gly Pro Ala Phe  
 340 345 350

Gln Met Val Cys Pro Ile Gly Trp Thr Gly Thr Val Ser Cys Thr Ser  
 355 360 365

Phe Asn Met Asp Thr Leu Ala Thr Thr Val Val Arg Thr Tyr Arg Arg  
 370 375 380

Ser Lys Pro Phe Pro His Arg Gln Gly Cys Ile Thr Gln Lys Asn Leu  
 385 390 395 400

Gly Glu Asp Leu His Asn Cys Ile Leu Gly Gly Asn Trp Thr Cys Val  
 405 410 415

Pro Gly Asp Gln Leu Leu Tyr Lys Gly Gly Ser Ile Glu Ser Cys Lys  
 420 425 430

Trp Cys Gly Tyr Gln Phe Lys Glu Ser Glu Gly Leu Pro His Tyr Pro  
 435 440 445

Ile Gly Lys Cys Lys Leu Glu Asn Glu Thr Gly Tyr Arg Leu Val Asp  
 450 455 460

Ser Thr Ser Cys Asn Arg Glu Gly Val Ala Ile Val Pro Gln Gly Thr  
 465 470 475 480

Leu Lys Cys Lys Ile Gly Lys Thr Thr Val Gln Val Ile Ala Met Asp  
 485 490 495

Thr Lys Leu Gly Pro Met Pro Cys Arg Pro Tyr Glu Ile Ile Ser Ser  
 500 505 510

Glu Gly Pro Val Glu Lys Thr Ala Cys Thr Phe Asn Tyr Thr Lys Thr  
 515 520 525

Leu Lys Asn Lys Tyr Phe Glu Pro Arg Asp Ser Tyr Phe Gln Gln Tyr  
 530 535 540

Met Leu Lys Gly Glu Tyr Gln Tyr Trp Phe Asp Leu Glu Val Thr Asp  
545 550 555 560

His His Arg Asp Tyr Phe Ala Glu Ser Ile Leu Val Val Val Ala  
565 570 575

Leu Leu Gly Gly Arg Tyr Val Leu Trp Leu Leu Val Thr Tyr Met Val  
580 585 590

Leu Ser Glu Gln Lys Ala Leu Gly  
595 600

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